

Original Article

# Serum transferrin as a liver fibrosis biomarker in patients with chronic hepatitis B

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**Background/Aims:** Transferrin and alpha-1 antitrypsin are reportedly associated with liver fibrosis. We evaluated the usefulness of serum transferrin and alpha-1 antitrypsin as new liver fibrosis markers in patients with chronic hepatitis B.

**Methods:** The study included 293 patients with chronic hepatitis B who underwent a liver biopsy between October 2005 and June 2009, and who had no history of hepatocellular carcinoma. Serum markers and liver fibrosis stages were compared.

**Results:** Univariate analysis revealed that age ( $P<0.001$ ), serum platelet count ( $P<0.001$ ), and serum alkaline phosphatase level ( $P=0.003$ ) differed significantly between the patients with and without liver cirrhosis. Serum transferrin levels were significantly lower in advanced fibrosis than in mild fibrosis in both univariate analysis ( $P=0.002$ ) and multivariate analysis ( $P=0.009$ ). In addition, the serum transferrin level was significantly lower in cirrhotic patients than in noncirrhotic patients ( $P=0.020$ ). However, the serum level of alpha-1 antitrypsin was not significantly associated with liver cirrhosis in patients with chronic hepatitis B.

**Conclusions:** Serum transferrin could be promising serum marker for predicting advanced liver fibrosis in patients with chronic hepatitis B. (*Clin Mol Hepatol* 2014;20:347-354)

**Keywords:** Chronic hepatitis B; Liver cirrhosis; Transferrin; Alpha-1 antitrypsin

## INTRODUCTION

Liver fibrosis involves the excessive deposition of extracellular matrix components in the liver as a consequence of chronic liver damage.<sup>1</sup> Accurate diagnosis of liver fibrosis is crucial to the management of patients with chronic hepatitis B (CHB) or chronic hepatitis C (CHC).<sup>2</sup>

Liver fibrosis is diagnosed by the histological analysis of liver biopsy specimens.<sup>3</sup> However, liver biopsy is invasive and is limited by

sampling errors, diagnostic inaccuracy, and hazards to the patient.<sup>4-6</sup> Therefore, there is a strong demand for reliable, organ-specific, noninvasive biomarkers of liver fibrosis to replace invasive needle biopsy.<sup>4</sup>

Serum-based tests of liver fibrosis have attracted more attention in recent years. Serum markers offer several advantages: they are much less invasive, analysis can be automated, and test performance is reproducible. The most widely published serum marker panel of liver fibrosis is the Fibrotest, which includes  $\alpha$ 2-

### Abbreviations:

AAT, alpha-1 antitrypsin; ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; AUC, area under the curve; CHB, chronic hepatitis B; GGT, gamma glutamyl transpeptidase; HBeAg, hepatitis B e antigen; PT, prothrombin time; ROC, receiver operating characteristics; Std, standard.

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Received : Aug. 13, 2014 / Revised : Sep. 1, 2014 / Accepted : Nov. 5, 2014

macroglobulin, apolipoprotein A1, haptoglobin, gamma glutamyl transpeptidase (GGT) and bilirubin.<sup>7-11</sup> Most serum-based panels have been generated from studies of patients with chronic hepatitis C.<sup>7,12-15</sup> On the contrary, in the case of CHB, similar models based on serum biochemical markers achieve a moderate correlation with liver fibrosis.<sup>16,17</sup> Therefore, there would be a need for finding accurate and specialized markers in CHB patients for predicting the degree of liver fibrosis.

Many previous studies suggested excessive iron accumulation in the hepatocyte accelerate hepatocellular damage, liver fibrosis, and eventually increase risk of hepatocellular carcinoma.<sup>18,19</sup> There were several studies to determine regulatory system of iron metabolism to maintain homeostasis and some studies proposed transferrin and hepcidin, which play a crucial role in iron metabolism, as surrogate markers for predicting severity of liver disease in the patients with chronic hepatitis C.<sup>20,21</sup> Recently, Xu et al. performed proteomic study to identify reliable non-invasive serum biomarkers of liver fibrosis in patients with CHB, and they reported decreased level of serum transferrin in cirrhotic patients compared with non-cirrhotic patients.<sup>22</sup> But there was no other study which validated serum transferrin as liver fibrosis marker in patients with CHB.

Alpha-1 antitrypsin (AAT) was known as most prominent protease inhibitor acting as anti-inflammatory cytokine and protecting the normal tissues during inflammation.<sup>23</sup> AAT was highly expressed in sera of patients with severe CHB and hepatocellular carcinoma compared with control.<sup>24</sup> There were many cytogenetic studies to find out the relationship of AAT and liver cirrhosis in patients with AAT deficiency.<sup>25,26</sup> However, there were very limited studies to investigate the association between serum AAT and liver fibrosis in viral hepatitis.<sup>27</sup>

Our previous pilot experiment for evaluating serum biomarker of liver fibrosis in CHB patients revealed that serum transferrin and AAT were significantly different between patients with advanced liver fibrosis and patients with mild fibrosis. In this study, we tried to investigate and validate the role of serum transferrin and AAT for predicting liver fibrosis in relatively large cohort of the patients with CHB.

## MATERIALS AND METHODS

### Patient selection

Between October 2005 and June 2009, 293 consecutive patients with CHB who underwent liver biopsies in six medical cen-

ters (Ajou University Hospital, Hallym University Chuncheon, CHA University Hospital, Inje University Busan Paik Hospital, Pusan National University Hospital, and Catholic University St. Vincent's Hospital) in South Korea were recruited in this study. The inclusion criteria were CHB patients who underwent a liver biopsy with no history of hepatocellular carcinoma and who did not receive antiviral treatment for 6 months prior to commencement of this study. Patients with other causes of liver disease or decompensated cirrhosis were not included.

Informed consent to participate in the study was obtained from all study subjects. The study protocols were approved by the Institutional Review Board of Human Research of Ajou University Hospital and the local Research Ethics Committees at all participating hospitals.

### Histologic examination

Liver biopsy specimens at least 10 mm long that contained at least 6 complete portal tracts were obtained from all patients. All biopsy specimens that met the inclusion criteria were staged by 2 central pathologists (YB Kim and YN Park). Hepatic fibrosis and necroinflammation were assessed using the Batts-Ludwig classification: F0=no fibrosis, F1=portal fibrosis without septa, F2=few septa, F3=many septa without cirrhosis, and F4=cirrhosis. Based on that system, grade 1 indicated necroinflammatory activities largely confined to the portal areas, grades 2-3 meant an extension beyond the portal areas, and grade 4 signified the bridging necrosis.

### Biochemical analysis

Serum samples were obtained and routine blood tests were performed at the time of liver biopsy and processed immediately. The serum biochemical parameters included total bilirubin, alanine aminotransferase, aspartate aminotransferase, GGT, alkaline phosphatase (ALP), albumin, blood urea nitrogen, creatinine, prothrombin time (PT), blood glucose, triglycerides, and total cholesterol levels. Transferrin and AAT level were measured with the Cobas Integra immunoturbidimetric assay (Roche, Basle, Switzerland).

### Statistics

The data about patients' characteristics are given as the mean±SD as appropriate. Comparisons between groups were performed by chi-square test, parametric independent *t* test, and

1-way analysis of variance test, as appropriate; differences between means were considered statistically significant for *P*-value of < 0.05. Logistic regression analysis was performed for multivariate analysis. Receiver operating characteristic (ROC) curve was used to evaluate the diagnostic accuracy of serum transferrin for discriminating patients with advanced fibrosis (F3 or F4) from without advanced fibrosis (F0 to F2). And area under the curve

(AUC) was calculated. All statistical tests were 2-sided and performed with SPSS ver. 18.0 (SPSS Inc., Chicago, IL, USA).

## RESULTS

### Patient characteristics

The baseline characteristics of the enrolled patients are depicted in Table 1. Among 293 patients, 205 patients (69.9%) were men; the mean age was 40.0 years (SD, 12.0 years; range, 14-70 years). Liver cirrhosis was present in 80 patients (27.3%). The distribution of the fibrosis stages was as follows: stage 0=12 patients (4.1%); stage 1=42 patients (14.3%); stage 2=82 patients (28.0%); stage 3=77 patients (26.3%); and stage 4=80 patients (27.3%). The grade of fibrosis was converted into a binomial variable of liver cirrhosis (F4, n=80) vs. no liver cirrhosis (F0, 1, 2, or 3, n=213) and mild fibrosis (F0, 1, 2, n=136) vs. advanced fibrosis (F3, F4, n=157).

Univariate analysis revealed that age (*P*<0.001), platelet counts (*P*<0.001), ALP (*P*=0.003) were significantly different between the patients with or without liver cirrhosis (Table 2).

### Measurement of serum transferrin and AAT levels

We evaluated the correlation of serum transferrin and AAT concentration with liver fibrosis in the cohort of CHB patients. We performed a one-way analysis of variance to compare means of serum

**Table 1.** Baseline characteristics of the patients (n=293)

Variables	
Age (years, [range])*	40.03±12.08 (14-70)
Gender (M/F [%])	205/88 (69.9/31.1)
Platelet (x10 <sup>3</sup> /mm <sup>3</sup> )*	186.29±61.00
Bilirubin (mg/dL)*	0.89±0.52
Aspartate aminotransferase (IU/L)*	89.68±100.69
Alanine aminotransferase (IU/L)*	124.31±147.34
Alkaline phosphatase (IU/L)*	161.41±91.46
GGT (IU/L)*	82.19±94.71
HBeAg positivity (n [%])	170 (58.0)
Stage of fibrosis (n [%])	
F0	12 (4.1)
F1	42 (14.3)
F2	82 (28.0)
F3	77 (26.3)
F4	80 (27.3)

\*Data expressed as mean ± SD.  
HBeAg, hepatitis B e antigen; GGT, gamma glutamyl transpeptidase.

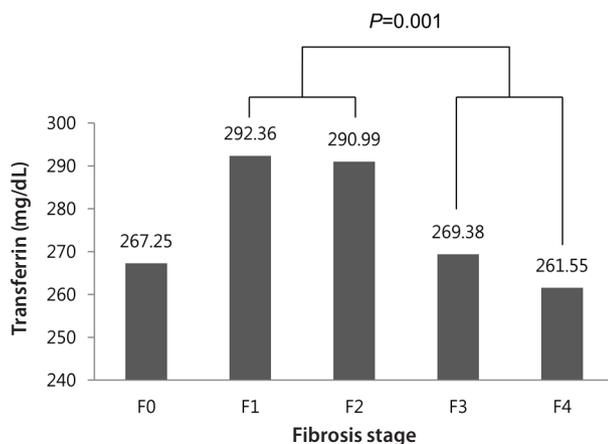
**Table 2.** Univariate and multivariate analysis for variables associated with liver cirrhosis

Variables*	No liver cirrhosis Stage ≤3 (n=213)	Liver cirrhosis Stage 4 (n=80)	<i>P</i> -value	
			Univariate	Multivariate
Age (years)	37.62±12.09	46.70±9.37	<0.001	0.003
Platelet (x10 <sup>3</sup> /mm <sup>3</sup> )	199.64±59.23	150.73±51.16	<0.001	<0.001
AST (IU/L)	91.24±91.93	84.04±121.87	0.590	
ALT (IU/L)	132.07±145.46	103.41±152.19	0.143	
GGT (IU/L)	74.94±70.72	107.06±148.87	0.141	
Albumin (g/dL)	4.19±0.46	4.08±0.60	0.086	0.125
Bilirubin (mg/dL)	0.85±0.52	0.97±0.49	0.066	
ALP (IU/L)	171.21±92.09	135.11±85.49	0.003	0.003
PT (INR)	1.08±0.14	1.12±0.15	0.061	
alpha-1 antitrypsin (mg/dL)	132.92±26.65	131.15±25.18	0.602	
Transferrin (mg/dL)	282.11±64.97	261.55±63.20	0.020	0.479

\*Data expressed as mean±standard deviation.  
AST, aspartate aminotransferase; ALT, alanine aminotransferase; GGT, gamma glutamyl transpeptidase; ALP, alkaline phosphatase; PT, prothrombin time.

transferrin levels among the fibrosis stages. Serum transferrin levels were significantly different between the groups ( $P=0.017$ ). Figure 1 shows the distribution of serum transferrin levels according to liver fibrosis stage. In the advanced fibrosis stage (F3 or F4), the serum transferrin level was significantly lower compared to patients with mild fibrosis stage (F1 or F2,  $P=0.001$ ).

Comparing between patients with liver cirrhosis (F4) and without cirrhosis (F0 to F3), serum platelet count ( $P<0.001$ ), serum ALP level ( $P=0.003$ ), age ( $P<0.001$ ) and serum transferrin level ( $P=0.020$ ) showed significant differences in univariate analysis.



**Figure 1.** Distribution of serum transferrin levels according to liver fibrosis stage. The serum transferrin level was significantly lower in advanced fibrosis (stage F3 or F4) than in mild fibrosis (stage F1 or F2).

Serum transferrin level was lower in cirrhotic patients when compared with non-cirrhotic patients. The variables of statistical significance ( $P<0.05$ ) on univariate analysis were entered into the multivariate analysis. In the multivariate analysis, serum platelet count ( $P<0.001$ ) and serum ALP level ( $P=0.003$ ) were still statistically significant, but serum transferrin was not significant. ( $P=0.479$ ). (Table 2)

Next, we analyzed the factors predicting advanced liver fibrosis. Serum transferrin level ( $P=0.002$ ), platelet count ( $P<0.001$ ), GGT ( $P=0.012$ ), albumin ( $P=0.004$ ), PT (INR) ( $P<0.001$ ) and patients' age ( $P<0.001$ ) showed significant differences in univariate analysis between patients with advanced liver fibrosis and without advanced fibrosis. In multivariate analysis, serum platelet count ( $P=0.001$ ) and serum transferrin level ( $P=0.009$ ) were revealed as independent factors for predicting advanced liver fibrosis. (Table 3)

Figure 2. shows the ROC curves of serum transferrin and serum platelet count to discriminate patients with advanced fibrosis from those without advanced fibrosis. The AUCs of serum transferrin and platelet count were 0.606 and 0.697, respectively ( $P=0.002$ ,  $P=0.000$ ).

Serum AAT levels were not significantly different between patients with and without liver cirrhosis (Table 2). No significant differences were observed in serum AAT levels among the fibrosis stages ( $P=0.582$ ).

Table 4 and Table 5. show results of univariate and multivariate analysis of transferrin as dependent variable and possible determinants, in order to verify transferrin as predicting marker of liver fi-

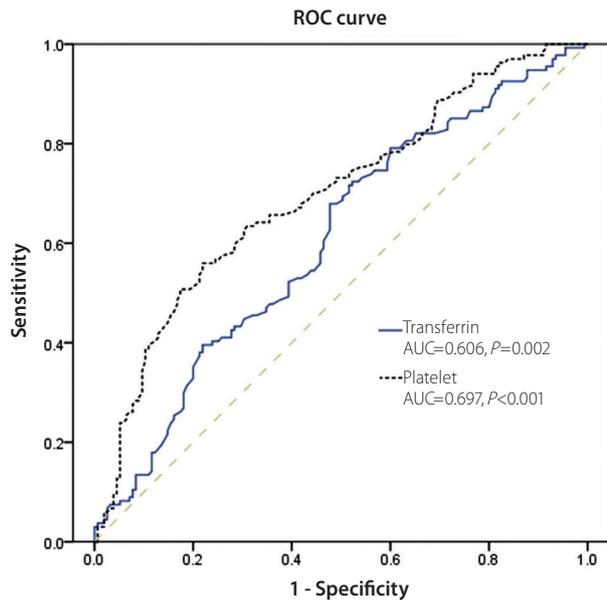
**Table 3.** Univariate and multivariate analysis for variables associated with advanced liver fibrosis

Variables*	No advanced liver fibrosis, Stage $\leq 2$ (n=136)	Advanced liver fibrosis, Stage 3 or 4 (n=157)	P-value	
			Univariate	Multivariate
Age (years)	36.18 $\pm$ 12.17	43.41 $\pm$ 10.98	<0.001	0.316
Platelet ( $\times 10^3/\text{mm}^3$ )	208.26 $\pm$ 59.02	167.29 $\pm$ 56.29	<0.001	0.001
AST (IU/L)	89.38 $\pm$ 99.85	89.95 $\pm$ 101.74	0.962	
ALT (IU/L)	137.07 $\pm$ 155.52	113.18 $\pm$ 139.36	0.167	
GGT (IU/L)	65.48 $\pm$ 62.87	97.07 $\pm$ 114.18	0.012	0.070
Albumin (g/dL)	4.26 $\pm$ 0.39	4.09 $\pm$ 0.57	0.004	0.153
Bilirubin (mg/dL)	0.84 $\pm$ 0.58	0.93 $\pm$ 0.45	0.170	
ALP (IU/L)	164.56 $\pm$ 85.01	158.68 $\pm$ 96.91	0.591	0.003
PT (INR)	1.06 $\pm$ 0.13	1.12 $\pm$ 0.15	<0.001	0.062
alpha-1 antitrypsin (mg/dL)	131.59 $\pm$ 23.25	133.19 $\pm$ 28.56	0.602	
Transferrin (mg/dL)	289.32 $\pm$ 66.76	265.39 $\pm$ 61.55	0.002	0.009

\*Data expressed as mean $\pm$ standard deviation.

AST, aspartate aminotransferase; ALT, alanine aminotransferase; GGT, gamma glutamyl transpeptidase; ALP, alkaline phosphatase; PT, prothrombin time.

bro sis. Serum ALP level ( $P=0.003$ ), serum albumin level ( $P<0.001$ ) and advanced liver fibrosis ( $P=0.011$ ) were statistically significant in both univariate and multivariate analysis.



**Figure 2.** ROC curves for the serum transferrin level and the serum platelet count for discriminating chronic hepatitis B patients with advanced liver fibrosis from those with mild fibrosis. The area under the curve was 0.606 for the serum transferrin level ( $P=0.002$ ) and 0.697 for the serum platelet count ( $P<0.001$ ). ROC, receiver operating characteristics; AUC, area under the curve.

## DISCUSSION

We performed the study to clarify the usefulness of serum transferrin and AAT levels in predicting liver fibrosis. Serum transferrin levels were decreased when mild fibrosis progressed to advanced fibrosis and cirrhosis, but serum AAT levels could not predict liver fibrosis in patients with CHB.

Hepatic iron deposition is well-known as leading cause of hepatic fibrosis and cirrhosis in hemochromatosis. Recently, hepatic iron deposition reported as important factor of hepatic fibrosis not only in the patients with hemochromatosis, but also in alcoholic liver disease, non-alcoholic steatohepatitis and chronic hepatitis C.<sup>28-30</sup> Several investigators have suggested a relationship between the severity of chronic hepatitis and the degree of liver iron deposition.<sup>21,31</sup>

Transferrin plays a crucial role in iron metabolism. Transferrin, produced by the hepatocytes, is a glycoprotein consisted with a single polypeptide chain and two iron-binding sites.<sup>32-34</sup> In our study, serum transferrin levels were elevated in the patients with mild fibrosis stage than those without hepatic fibrosis. However, it decreased when mild fibrosis progressed to advanced fibrosis and cirrhosis. Several reports on serum transferrin levels and liver fibrosis are comparable to ours. Potter et al. reported that transferrin level was increased in patients with alcoholic fatty livers but significantly decreased in cirrhotic patients.<sup>35</sup> Ibrahim et al. reported that serum transferrin level was slightly increased in a pediatric liver disease group without liver cirrhosis, but was decreased in

**Table 4.** Univariate analysis for variables associated with serum transferrin

Variables	Unstandardized coefficients		Standardized coefficients	<i>t</i>	<i>P</i> -value
	B	Std. error	Beta		
Age (years)	-5.163	3.038	-0.099	-1.699	0.090
Hemoglobin (g/dL)	-1.988	2.279	-0.051	-0.872	0.384
AST (IU/L)	0.033	0.038	0.051	0.861	0.390
ALT (IU/L)	0.049	0.026	0.111	1.907	0.058
Bilirubin (mg/dL)	-2.369	7.456	-0.019	-0.318	0.751
ALP (IU/L)	0.092	0.042	0.129	2.169	0.031
GGT (IU/L)	0.027	0.048	0.037	0.557	0.578
Albumin (g/dL)	26.580	7.570	0.205	3.511	0.001
PT (INR)	-39.435	28.016	-0.085	-1.408	0.160
HBV DNA (IU/mL)	8.139	6.177	0.083	1.318	0.189
Fibrosis(F0,1,2 vs. F3,4)	-23.928	7.499	-0.184	-3.191	0.002

Std., standard; AST, aspartate aminotransferase; ALT, alanine aminotransferase; GGT, gamma glutamyl transpeptidase; ALP, alkaline phosphatase; PT, prothrombin time.

**Table 5.** Multivariate analysis for variables associated with serum transferrin

	Unstandardized coefficients		Standardized coefficients	<i>t</i>	<i>P</i> -value
	B	Std. error	Beta		
ALP (IU/L)	0.125	0.042	0.175	2.986	0.003
Albumin (g/dL)	31.025	8.203	0.225	3.782	0.000
Fibrosis (F0,1,2 vs. F 3,4)	-19.386	7.578	-0.148	-2.558	0.011

Std., standard; ALP, alkaline phosphatase.

cirrhosis patients.<sup>36</sup> These reports suggested that transferrin synthesis appeared to be raised during the early stage of liver disease, and thereafter it was decreased in advanced liver fibrosis, probably reflecting diminished synthetic capacity by the liver because of severe liver cell injury. Our study revealed that serum albumin level is also independent predictor of transferrin. This result suggests the possibility of an additional mechanism of transferrin suppression in advanced fibrosis other than decreased synthetic function. However, the exact pathophysiologic mechanism has not been clearly proven yet. Further basic research for identifying the mechanism will be needed.

There were several researches about iron metabolism alteration in chronic liver disease. Hcpidin plays a critical role in the maintenance of iron homeostasis. Several investigators have reported that hepcidin expression of hepatocyte was decreased at chronic hepatitis. Decreased hepcidin expression resulted in appearance of non-transferrin-bound iron, which known as a potentially toxic iron form, and to the development of parenchymal iron overload. Hcpidin expression is regulated by several factors including serum transferrin saturation.<sup>37,38</sup> Considering results of our study, hepatic transferrin secretion might be increased to down regulate transferrin saturation due to compensate decreased hepcidin expression at mild fibrosis stage.

AAT is a protease inhibitor belonging to the serpin superfamily. It is an acute phase reactant, and its levels rise following tissue injury and inflammation.<sup>39,40</sup> In our previous study, AAT level was significantly higher in patients with significant inflammation in univariate analysis but not in multivariate analysis.<sup>41</sup> In current study, serum AAT was not associated with liver fibrosis in patients with CHB. Considering the function of AAT, it appeared to be more correlated with hepatic necroinflammatory activity than liver fibrosis. Further research will be needed to evaluate the clinical usefulness of AAT for predicting hepatic inflammation and fibrosis in patients with chronic hepatitis.

Our study had some limitations. First, our study did not separate HBeAg-positive patients from HBeAg-negative patients. The dif-

ferences in clinical course or outcome in these two groups have been demonstrated in previous studies.<sup>16,42</sup> Second, we didn't investigate other serum markers of iron metabolism like iron, ferritin, and haptoglobin which could affect the serum transferrin level. Third, our study did not clarify the relationship between serum transferrin level and hepatic iron deposition.

In conclusion, we showed that serum transferrin could be promising liver fibrosis marker in patients with CHB. Serum transferrin level was lower in the advanced fibrosis stage compared with mild fibrosis stage in the patients with CHB. Further study will be needed to establish the usefulness of serum transferrin level as potent liver fibrosis marker in patients with viral hepatitis. To our knowledge, this is the first validation study of serum transferrin level as a liver fibrosis-predicting marker in a relatively large number of CHB patients.

### Acknowledgement

This work was supported by a grant from the Ministry of Health and Welfare, Republic of Korea (no. A102065)

### Conflicts of Interest

The authors have no conflicts to disclose.

### REFERENCES

1. Friedman SL. Liver fibrosis -- from bench to bedside. *J Hepatol* 2003;38 (Suppl 1):S38-S53.
2. Suk KT, Baik SK, Yoon JH, Cheong JY, Paik YH, Lee CH, et al. Revision and update on clinical practice guideline for liver cirrhosis. *Korean J Hepatol* 2012;18:1-21.
3. Bravo AA, Sheth SG, Chopra S. Liver biopsy. *N Engl J Med* 2001;344:495-500.
4. Abdi W, Millan JC, Mezey E. Sampling variability on percutaneous liver biopsy. *Arch Intern Med* 1979;139:667-669.

5. Terjung B, Lemnitzer I, Dumoulin FL, Effenberger W, Brackmann HH, Sauerbruch T, et al. Bleeding complications after percutaneous liver biopsy. An analysis of risk factors. *Digestion* 2003;67:138-145.
6. Bedossa P, Dargere D, Paradis V. Sampling variability of liver fibrosis in chronic hepatitis C. *Hepatology* 2003;38:1449-1457.
7. Imbert-Bismut F, Ratziu V, Pieroni L, Charlotte F, Benhamou Y, Poynard T, et al. Biochemical markers of liver fibrosis in patients with hepatitis C virus infection: a prospective study. *Lancet* 2001;357:1069-1075.
8. Rossi E, Adams L, Prins A, Bulsara M, de Boer B, Garas G, et al. Validation of the fibrotest biochemical markers score in assessing liver fibrosis in hepatitis C patients. *Clin Chem* 2003;49:450-454.
9. Poynard T, McHutchison J, Manns M, Myers RP, Albrecht J. Biochemical surrogate markers of liver fibrosis and activity in a randomized trial of peginterferon alfa-2b and ribavirin. *Hepatology* 2003;38:481-492.
10. Le Calvez S, Thabut D, Messous D, Munteanu M, Ratziu V, Imbert-Bismut F, et al. The predictive value of fibrotest vs. APRI for the diagnosis of fibrosis in chronic hepatitis C. *Hepatology* 2004;39:862-863; author reply 863.
11. Thabut D, Simon M, Myers RP, Messous D, Thibault V, Imbert-Bismut F, et al. Noninvasive prediction of fibrosis in patients with chronic hepatitis C. *Hepatology* 2003;37:1220-1221; author reply 1221.
12. Fornis X, Ampurdanes S, Llovet JM, Aponte J, Quinto L, Martinez-Bauer E, et al. Identification of chronic hepatitis C patients without hepatic fibrosis by a simple predictive model. *Hepatology* 2002;36:986-992.
13. Wai CT, Greenson JK, Fontana RJ, Kalbfleisch JD, Marrero JA, Conjeevaram HS, et al. A simple noninvasive index can predict both significant fibrosis and cirrhosis in patients with chronic hepatitis C. *Hepatology* 2003;38:518-526.
14. Lok AS, Ghany MG, Goodman ZD, Wright EC, Everson GT, Sterling RK, et al. Predicting cirrhosis in patients with hepatitis C based on standard laboratory tests: results of the HALT-C cohort. *Hepatology* 2005;42:282-292.
15. Leroy V, Monier F, Bottari S, Trocme C, Sturm N, Hilleret MN, et al. Circulating matrix metalloproteinases 1, 2, 9 and their inhibitors TIMP-1 and TIMP-2 as serum markers of liver fibrosis in patients with chronic hepatitis C: comparison with PIINP and hyaluronic acid. *Am J Gastroenterol* 2004;99:271-279.
16. Myers RP, Tainturier MH, Ratziu V, Piton A, Thibault V, Imbert-Bismut F, et al. Prediction of liver histological lesions with biochemical markers in patients with chronic hepatitis B. *J Hepatol* 2003;39:222-230.
17. Cheong JY, Um SH, Seo YS, Kim DJ, Hwang SG, Lee YJ, et al. Non-invasive index for predicting significant liver fibrosis: comparison of diagnostic performances in patients with chronic hepatitis B and C. *Dig Dis Sci* 2011;56:555-563.
18. Franchini M, Targher G, Capra F, Montagnana M, Lippi G. The effect of iron depletion on chronic hepatitis C virus infection. *Hepatol Int* 2008;2:335-340.
19. Tirnitz-Parker JE, Glanfield A, Olynyk JK, Ramm GA. Iron and hepatic carcinogenesis. *Crit Rev Oncog* 2013;18:391-407.
20. Vagu C, Sultana C, Ruta S. Serum iron markers in patients with chronic hepatitis C infection. *Hepat Mon* 2013;13:e13136.
21. Fletcher LM, Halliday JW, Powell LW. Interrelationships of alcohol and iron in liver disease with particular reference to the iron-binding proteins, ferritin and transferrin. *J Gastroenterol Hepatol* 1999;14:202-214.
22. Xu MY, Qu Y, Jia XF, Wang ML, Liu H, Wang XP, et al. Serum proteomic MRM identify peptide ions of transferrin as new fibrosis markers in chronic hepatitis B. *Biomed Pharmacother* 2013;67:561-567.
23. Van Molle W, Denecker G, Rodriguez I, Brouckaert P, Vandenaabeele P, Libert C. Activation of caspases in lethal experimental hepatitis and prevention by acute phase proteins. *J Immunol* 1999;163:5235-5241.
24. Tan XF, Wu SS, Li SP, Chen Z, Chen F. Alpha-1 antitrypsin is a potential biomarker for hepatitis B. *Virology* 2011;8:274.
25. Pferdmeiges DC, Baumann U, Müller-Heine A, Framke T, Pfister ED. Prognostic marker for liver disease due to alpha1-antitrypsin deficiency. *Klin Padiatr* 2013;225:257-262.
26. Kohnlein T, Rifai K. [Alpha1-antitrypsin deficiency]. *Internist (Berl)* 2010;51 (Suppl 1):S269-S276.
27. Beloborodova EV, Beloborodova EI, Akbasheva OE, Serebrov Vlu, Chernogoriuk GE, Rachkovskii MI, et al. [The parameters of collagen proteolytic and metabolic systems in chronic liver diseases of viral and toxic etiologies]. *Ter Arkh* 2010;82:29-34.
28. Metwally MA, Zein CO, Zein NN. Clinical significance of hepatic iron deposition and serum iron values in patients with chronic hepatitis C infection. *Am J Gastroenterol* 2004;99:286-291.
29. Marchesini G, Brizi M, Bianchi G, Tomassetti S, Bugianesi E, Lenzi M, et al. Nonalcoholic fatty liver disease a feature of the metabolic syndrome. *Diabetes* 2001;50:1844-1850.
30. Bassett ML, Halliday JW, Powell LW. Value of hepatic iron measurements in early hemochromatosis and determination of the critical iron level associated with fibrosis. *Hepatology* 1986;6:24-29.
31. Fletcher LM. Alcohol and iron: one glass of red or more? *J Gastroenterol Hepatol* 1996;11:1039-1041.
32. Wang J, Pantopoulos K. Regulation of cellular iron metabolism. *Biochem. J* 2011;434:365-381.
33. Andrews NC. Disorders of iron metabolism. *N Engl J Med* 1999;341:1986-1995.
34. de Jong G, van Dijk JP, van Eijk HG. The biology of transferrin. *Clin Chim Acta.* 1990;190:1-46.
35. Potter BJ, Chapman RW, Nunes RM, Sorrentino D, Sherlock S. Transferrin metabolism in alcoholic liver disease. *Hepatology* 1985;5:714-721.
36. Ibrahim AM, Ali MM, Ghanem HM, Mousa AM, Ghafar TYA. Simple and rapid detection of liver cirrhosis in children by tracking serum IgA/transferrin ratio. *J Exp Integr Med* 2011;1:117-121.
37. Loréal O, Cavey T, Bardou-Jacquet E, Guggenbuhl P, Ropert M, Brissot P. Iron, hepcidin and the metal connection. *Front Pharmacol.* 2014;5:128.
38. Hino K, Nishina S, Hara Y. Iron metabolic disorder in chronic hepatitis

- C: Mechanisms and relevance to hepatocarcinogenesis. *J Gastroenterol Hepatol.* 2013;28:93-98.
39. Travis J, Salvesen GS. Human plasma proteinase inhibitors. *Annu Rev Biochem* 1983;52:655-709.
40. Dickson I, Alper CA. Changes in serum proteinase inhibitor levels following bone surgery. *Clin Chim Acta* 1974;54:381-385.
41. Cho HJ, Kim SS, Ahn SJ, Bae CB, Kim HG, Kim YJ, et al. Serum markers for predicting significant necroinflammatory activity in patients with chronic hepatitis B. *Clin Biochem* 2012;45:1564-1567.
42. Hui AY, Chan HL, Wong VW, Liew CT, Chim AM, Chan FK, et al. Identification of chronic hepatitis B patients without significant liver fibrosis by a simple noninvasive predictive model. *Am J Gastroenterol* 2005;100:616-623.
41. Cho HJ, Kim SS, Ahn SJ, Bae CB, Kim HG, Kim YJ, et al. Serum markers