



Endocrinol Metab 2016;31:134-141 http://dx.doi.org/10.3803/EnM.2016.31.1.134 pISSN 2093-596X · eISSN 2093-5978

Glucose-Dependent Insulinotropic Peptide Level Is Associated with the Development of Type 2 Diabetes Mellitus

Original

Article

Sunghwan Suh^{1,*}, Mi Yeon Kim^{2,*}, Soo Kyoung Kim³, Kyu Yeon Hur², Mi Kyoung Park¹, Duk Kyu Kim¹, Nam H. Cho⁴, Moon-Kyu Lee²

¹Division of Endocrinology and Metabolism, Department of Internal Medicine, Dong-A Medical Center, Dong-A University College of Medicine, Busan; ²Division of Endocrinology and Metabolism, Department of Medicine, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul; ³Division of Endocrinology and Metabolism, Department of Internal Medicine, Gyeongsang National University School of Medicine, Jinju; ⁴Department of Preventive Medicine, Ajou University School of Medicine, Suwon, Korea

Background: Incretin hormone levels as a predictor of type 2 diabetes mellitus have not been fully investigated. Therefore, we measured incretin hormone levels to examine the relationship between circulating incretin hormones, diabetes, and future diabetes development in this study.

Methods: A nested case-control study was conducted in a Korean cohort. The study included the following two groups: the control group (n=149), the incident diabetes group (n=65). Fasting total glucagon-like peptide-1 (GLP-1) and total glucose-dependent insulinotropic peptide (GIP) levels were measured and compared between these groups.

Results: Fasting total GIP levels were higher in the incident diabetes group than in the control group $(32.64\pm22.68 \text{ pmol/L vs.} 25.54\pm18.37 \text{ pmol/L}, P=0.034)$. There was no statistically significant difference in fasting total GLP-1 levels between groups $(1.14\pm1.43 \text{ pmol/L vs.} 1.39\pm2.13 \text{ pmol/L}, P=0.199)$. In multivariate analysis, fasting total GIP levels were associated with an increased risk of diabetes (odds ratio, 1.005; P=0.012) independent of other risk factors.

Conclusion: Fasting total GIP levels may be a risk factor for the development of type 2 diabetes mellitus. This association persisted even after adjusting for other metabolic parameters such as elevated fasting glucose, hemoglobin A1c, and obesity in the pre-diabetic period.

Keywords: Gastric inhibitory polypeptide; Diabetes mellitus, type 2; Glucagon-like peptide 1

Received: 13 August 2015, Revised: 4 September 2015,
Accepted: 15 September 2015
Corresponding authors: Nam H. Cho
Department of Preventive Medicine, Ajou University School of Medicine,
164 World cup-ro, Yeongtong-gu, Suwon 16499, Korea
Tel: +82-31-219-4217, Fax: +82-31-219-5084, E-mail: chnaha@ajou.ac.kr
Moon-Kyu Lee
Division of Endocrinology and Metabolism, Department of Medicine, Samsung
Medical Center, Sungkyunkwan University School of Medicine, 81 Irwon-ro,
Gangnam-gu, Seoul 06351, Korea

Tel: +82-2-3410-3431, **Fax:** +82-2-3410-0393, **E-mail:** leemk@skku.edu *These authors contributed equally to this work.

Copyright © 2016 Korean Endocrine Society

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/ licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

EnM

INTRODUCTION

Glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic peptide (GIP) are incretin hormones that potentiate glucose-stimulated insulin secretion from β -cells [1]. Recently, new therapeutic agents such as GLP-1 analogs and dipeptidyl peptidase-4 (DPP4) inhibitors were introduced to clinical practice with proven efficacy of glucose control [1]. Development of these novel medications was based on a marked reduction of incretin effects in patients with type 2 diabetes mellitus (T2DM) [2]. In patients with T2DM, the reduction of incretin effects was largely due to decreased GLP-1 secretion after nutrient stimulation or ineffective GIP action [3]. Nevertheless, it has been demonstrated recently that the incretin effect is not impaired in Japanese and Korean T2DM subjects [4,5]. Recent meta-analyses of clinical studies also suggested that patients with T2DM, in general, do not exhibit reduced GLP-1 secretion in response to an oral glucose tolerance test (OGTT) or meal test [6]. There are ethnic differences in the pathogenesis of T2DM, especially in Caucasian and East Asians [7]. However, studies of incretin action in East Asians are limited.

It is well established that the risk of diabetes in prediabetic subjects is much higher compared to those with normal glucose tolerance (NGT) [8]. Even among subjects with NGT, an upper normal level of fasting plasma glucose (FPG) is a predictor of T2DM [9]. However, incretin hormone levels as a predictor of T2DM have not been fully investigated because incretin levels have only been measured in small groups of subjects [10,11] due to the relatively laborious and time-consuming nature of the laboratory methods associated with this measurement. The present study was conducted to evaluate incretin hormone levels before diabetes develops and to determine the role of incretin hormones as predictors of diabetes development.

METHODS

Study design and participants

A nested case-control analysis was performed using participants of an Ansung cohort study. The design and baseline characteristics of the Ansung-Ansan cohort study have been described in detail elsewhere [12]. Briefly, it is an ongoing prospective community-based epidemiological study that is a part of the Korean Health and Genome Study which was conducted to investigate trends in diabetes and associated risk factors. The baseline examination was performed in 2001 to 2002, and biennial followup examinations were continued through 2012. Of the 5,018 subjects who were surveyed in Ansung, complete data from the baseline investigation and frozen samples for further analysis were available for 1,371 participants who registered for the cohort study in the first year (2001). Among the 1,371 subjects, the results of OGTT were available for 1,358 subjects. During both initial screening and follow-up visit, the definitions of NGT, prediabetes and diabetes were based on plasma glucose levels during the 75 g OGTT according to the 1997 American Diabetes Association criteria. NGT was defined as FPG level <6.1 mmol/ L and 2-hour plasma glucose <7.8 mmol/L. Prediabetes was defined as 6.1 mmol/L \leq FPG level < 7.0 mmol/L or 7.8 mmol/L \leq 2-hour plasma glucose<11.1 mmol/L. Diabetes was defined as FPG concentration \geq 7.0 mmol/L or 2-hour plasma glucose \geq 11.1 mmol/L or current treatment with oral antidiabetic drugs or insulin [13]. At the time of initial screening in 2001, there were 948 subjects with NGT, and 261 subjects with prediabetes. Among the subjects who showed NGT in 2001, 23.9% (227/948) developed impaired fasting glucose or impaired glucose tolerance (IGT) and 6.9% (65/948) developed diabetes by 2009. The "incident diabetes group (NGT→diabetes mellitus group)" in the present study consisted of the 65 subjects who developed diabetes during the follow-up period. Among the 656 subjects who maintained NGT during the 8-year follow-up period, 149 subjects were randomly selected and matched to diabetic subjects by sex and age. These 149 subjects were defined as the "control group (NGT→NGT group)" (Fig. 1). Circulating levels of total GLP-1 and total GIP were measured in selected samples to examine the relationship between basal incretin hormones and diabetes development. The study protocol was approved by the Institutional Review Boards of the Samsung Medical Center (Approval number 2010-11-068-001).

Data collection and biochemical analyses

Anthropometric parameters and blood pressure were measured using standard methods. FPG, insulin, total cholesterol, triglycerides, and high density lipoprotein cholesterol were measured, and the results were obtained from the main database. Homeostasis model of assessment-insulin resistance (HOMA-IR) was defined as [fasting insulin (μ U/mL)×fasting glucose (mmol/L)]/22.5. Homeostasis model of assessment- β cell function (HOMA-B) was calculated using (20×fasting insulin in μ U/mL)/(fasting glucose in mmol/L–3.5) [14]. Total GLP-1 and total GIP were measured in the stored samples. The plasma concentrations of total GLP-1 (ALPCO Diagnostics, Windham, NH, USA) and total GIP (Millipore Corp., Bedford, MA, USA) were measured without an ethanol extraction step by an enzyme-linked immunosorEnM



Fig. 1. Flow diagram of the study. OGTT, oral glucose tolerance test.

bent assay. Samples were frozen at -70° C and never thawed until they were moved to Samsung Medical Center for analyses. The biochemical analysis of incretin hormones was performed in duplicate by a single trained technician in the Department of Laboratory Medicine and Genetics at the Samsung Medical Center.

Statistical analyses

Statistical analyses to compare baseline characteristics between and among the groups were conducted using one-way analysis of variance for normally distributed data. The Kruskal-Wallis test was used for analysis of skewed data. Categorical variables were compared using Pearson and chi-square test. Univariate analyses were performed to assess the association between diabetes development and clinical parameters. Multivariate analysis was performed using variables from the univariate analyses that were significant at P<0.05. A binary logistic regression model was used for univariate and multivariate analyses. P<0.05 was considered statistically significant. A binary logistic regression model was used for development of diabetes according to incretin quintiles. Statistical analyses were performed using IBM SPSS version 19 (IBM Co., Armonk, NY, USA).

RESULTS

Descriptive data

Demographic characteristics and laboratory results are summa-

136 www.e-enm.org

rized in Table 1. Demographic characteristics such as age and sex were not significantly different between groups. On the other hand, significant differences in waist circumference, body mass index (BMI), systolic blood pressure, diastolic blood pressure, FPG, postprandial glucose, hemoglobin A1c (HbA1c), triglyceride, aspartate transaminase, alanine transaminase (ALT), γ -glutamyl transpeptidase were observed. Those discrepancies were primarily observed in subjects who had diabetes upon initial enrollment. To investigate the role of incretin hormones as predictors of incident diabetes, total GLP-1 and total GIP levels were compared between patients who developed diabetes and those who maintained NGT. Total GLP-1 levels were not significantly different between the two groups who developed diabetes and those who maintained NGT (P=0.199). Nonetheless, there were significant differences in total GIP levels between patients who developed diabetes and those who did not (P=0.034). Correlations between incretin hormones and HOMA-B and HOMA-IR were not significant (data not shown).

Risk estimates for diabetes development

Parameters that were significantly associated with diabetes development included waist circumference, BMI, blood pressure, FPG, postprandial glucose, HbA1c, triglycerides, family history of diabetes, and total GIP levels (Table 2). In multivariate analyses, incretin levels were not significantly associated with the development of diabetes (model 1 of Table 2). After select-

Characteristic	$NCT \rightarrow NCT (n = 140)$	$NCT \rightarrow DM(n=65)$	Ducha
	$\frac{1}{2} \frac{1}{2} \frac{1}$	$\frac{1}{50(405(5))}$	
Age, yr	62 (54-65)	35 (49.5-65)	0.144
Sex, male:Temale	66:83	35:30	0.347
Waist circumference, cm	83.22±8.37	86.85±8.04	0.003
BMI, kg/m ²	23.09 (21.67–24.97)	24.45 (22.13–26.51)	0.040
SBP, mm Hg	118.67 (108.33–127.67)	129.33 (114.33–139.33)	< 0.001
DBP, mm Hg	71.33 (67.33–77.67)	80.00 (72.33-85.33)	< 0.001
Fasting glucose, mmol/L	4.28 (4.08-4.51)	4.63 (4.25–5.09)	< 0.001
Fasting insulin, pmol/L	39.24 (31.00–79.00)	40.28 (43.50-87.00)	0.370
Glucose, PP2, mmol/L ^a	5.69 (4.73-6.57)	6.10 (5.51–7.09)	0.001
Insulin, PP2, pmol/L ^a	127.09 (107.25-345.50)	154.18 (126.50-329.00)	0.517
Hemoglobin A1c, %	5.30 (5.09–5.50)	5.63 (5.40-5.91)	< 0.001
Total cholesterol, mmol/L	4.59 ± 0.68	4.68±0.90	0.450
Triglyceride, mmol/L	1.43 (1.12–1.89)	1.73 (1.34–2.31)	0.033
LDL-C, mmol/L	2.70 (2.30-4.32)	2.73 (2.22–3.39)	0.799
HDL-C, mmol/L	1.13 (0.98–1.29)	1.08 (0.96–1.78)	0.085
Albumin, g/L	40.6 (39.13-41.37)	40.6 (39.50-41.37)	0.144
AST, U/L	25.62 (22.65–30.57)	27.60 (23.64–33.55)	0.004
ALT, U/L	21.42 (17.18–26.72)	23.54 (19.30-33.60)	< 0.001
γ-GTP, U/L	13.08 (10.07–23.12)	22.12 (14.59-43.20)	< 0.001
HOMA-B	141.17 (84.67–213.49)	115.38 (68.98–176.72)	0.412
HOMA-IR	1.11 (0.58–1.52)	1.18 (0.79–1.81)	0.223
Total GLP-1, pmol/L	1.39 (0.69–2.82)	1.14 (0.65–2.08)	0.199
Total GIP, pmol/L	25.54 (7.17-43.91)	32.64 (9.96–55.32)	0.034
Family history of diabetes, yes:no	3:146	7:58	< 0.001

Values are expressed as median (interquartile range) or mean \pm SD.

NGT, normal glucose tolerance; DM, diabetes mellitus; BMI, body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure; PP2, postprandial 2 hours; LDL-C, low density lipoprotein cholesterol; HDL-C, high density lipoprotein cholesterol; AST, aspartate transaminase; ALT, alanine transaminase; γ -GTP, gamma-glutamyl transpeptidase; HOMA-B, homeostasis model of assessment- β cell function; HOMA-IR, homeostasis model of assessment-insulin resistance; GLP-1, glucagon-like peptide-1; GIP, glucose-dependent insulinotropic peptide.

ing the most relevant variables to predict diabetes occurrence, a second model was established (model 2 of Table 2). In this second model, fasting total GIP levels were significantly associated with diabetes development, whereas GLP-1 levels were not. In both univariate and multivariate analyses, higher HbA1c was the most significant predictor of diabetes development. In multivariable logistic regression analyses adjusted for age, sex and BMI, total GIP levels in the fifth quintile were associated with higher odds ratios of future diabetes development, while total GLP-1 levels were not. This association remained significant after additional adjustment for HbA1c (Table 3).

DISCUSSION

The purpose of this study was to investigate the association between incretin hormones and diabetes development in a community-based cohort. The novel finding is that fasting GIP levels were elevated before diabetes development even in subjects with NGT. In patients with T2DM, the reduced effects of incretin are well elucidated [2]. Reduced incretin effects have been explained by diminished GLP-1 levels in plasma and deteriorated GIP action [3]. Although impaired GLP-1 secretion after nutritional stimuli in diabetic patients has been reported in several papers [15,16], GLP-1 secretion is currently believed to be preserved in diabetic patients [6]. Some researchers have pro-

Table 2. Relationship between Development of Diabetes and Clinical Parameters							
Variable	Univariate analysis	Model 1	Model 2				
Age, yr	0.974 (0.941–1.009)						
Female sex	1.467 (0.817–2.634)						
Waist circumference, cm	1.055 (1.017–1.095)	1.093 (1.001–1.193)	1.046 (0.999–1.095)				
BMI, kg/m ²	1.114 (1.004–1.236)	0.812 (0.630-1.036)					
SBP, mm Hg	1.041 (1.020–1.062)	0.995 (0.959–1.033)					
DBP, mm Hg	1.101 (1.060–1.143)	1.104 (1.033–1.179)	1.096 (1.049–1.145)				
Fasting glucose, mmol/L	1.106 (1.060–1.154)	1.114 (1.035–1.198)	1.104 (1.050-1.162)				
Fasting insulin, pmol/L	1.014 (0.982–1.047)						
Glucose, PP2, mmol/L ^a	1.026 (1.010-1.042)	1.016 (0.996–1.036)					
Insulin, PP2, pmol/L ^a	1.005 (0.990-1.019)						
Hemoglobin A1c, %	6.605 (2.842–15.347)	5.314 (1.870–15.098)	4.662 (1.789–12.145)				
Total cholesterol, mmol/L	1.004 (0.994–1.014)						
Triglyceride, mmol/L	1.004 (1.000-1.007)	1.002 (0.997-1.007)					
LDL-C, mmol/L	1.001 (0.991–1.011)						
HDL-C, mmol/L	0.972 (0.940-1.006)						
Albumin, g/L	3.823 (0.631–23.182)						
HOMA-B	0.999 (0.998-1.001)						
HOMA-IR	1.095 (0.937-1.280)						
Total GLP-1, pmol/L	0.936 (0.845-1.037)	0.954 (0.839–1.086)	0.941 (0.833-1.063)				
Total GIP, pmol/L	1.004 (1.000–1.007)	1.003 (0.998-1.008)	1.005 (1.000-1.010)				
Family history of DM, yes/no	5.874 (1.468–23.494)	2.110 (0.330-13.472)					

Values are expressed as odds ratio (95% confidence interval).

BMI, body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure; PP2, postprandial 2 hours; LDL-C, low density lipoprotein cholesterol; HDL-C, high density lipoprotein cholesterol; HOMA-B, homeostasis model of assessment-β cell function; HOMA-IR, homeostasis model of assessment-insulin resistance; GLP-1, glucagon-like peptide-1; GIP, glucose-dependent insulinotropic peptide; DM, diabetes mellitus. ^aAccording to the method section, it should be 2-hour plasma glucose during 75 g oral glucose tolerance test. Or, the authors should mention that they performed postprandial measures, too.

	1st	2nd	3rd	4th	5th
GLP-1 quintile					
Model 1 ^a	1.00	1.34 (0.56-3.21)	1.11 (0.45-2.77)	0.94 (0.36-2.45)	0.65 (0.24–1.74)
Model 2 ^b	1.00	1.57 (0.64–3.87)	1.41 (0.54–3.65)	1.22 (0.45-3.32)	0.89 (0.32-2.52)
Model 3 ^c	1.00	2.05 (0.76-5.55)	1.80 (0.64-5.08)	1.38 (0.47-4.01)	0.79 (0.26-2.41)
GIP quintile					
Model 1 ^a	1.00	1.51 (0.54-4.24)	1.84 (0.67–5.05)	2.34 (0.88-6.26)	$2.89(1.08-7.75)^d$
Model 2 ^b	1.00	1.53 (0.52-4.45)	2.13 (0.75-6.06)	2.74 (0.99–7.55)	3.64 (1.30-10.20) ^d
Model 3°	1.00	1.60 (0.52-4.94)	2.31 (0.76-6.99)	2.52 (0.87-7.31)	3.82 (1.26-11.54) ^d

Values are expressed as odds ratio (95% confidence interval).

GLP-1, glucagon-like peptide-1; GIP, glucose-dependent insulinotropic peptide.

^aModel 1: unadjusted; ^bModel 2: age, sex, body mass index (BMI) adjusted; ^cModel 3: age, sex, BMI, hemoglobin A1c adjusted; ^dP<0.05.

posed that reduced incretin effect is a consequence of diabetic status, as deterioration in glucose homeostasis can develop in the absence of any impairment in GIP or GLP-1 levels [10].

The findings of a large cohort study demonstrated that fasting and integrated postprandial concentrations of total and active GLP-1 are not significantly different between subjects with NGT and those with IGT [17]. In addition, recent case-control study also found no difference in fasting GLP-1 among normal, prediabetes and diabetes groups [18]. The present study had similar findings when comparing GLP-1 levels in normal and diabetic subjects on enrollment (data not shown).

GIP was the first incretin identified. In addition to stimulating insulin secretion, GIP plays regulatory roles in the maintenance, growth and survival of pancreatic islets, as well as impacting on adipocyte function [19]. In the present study, GIP levels were increased significantly in subjects who later developed diabetes. This result is also in line with previous report showing that GIP levels were significantly elevated in newly-diagnosed diabetes group when compared with the normal group [18]. The mechanisms for these findings are unclear, but GIP elevations might be caused by GIP receptor resistance or defective GIP receptor expression. GIP receptor gene polymorphism was shown to be associated with diabetes and the metabolic syndrome, and GIP receptor polymorphism has begun to receive more attention than GIP plasma levels [20]. GIP/GIP receptor axis is also known to be disrupted in insulin-resistant states, such as obesity [21]. A meta-analysis of nine genome-wide association studies was conducted in order to identify T2DM-associated loci, and it was concluded that genetic variation in the GIP receptor influences glucose and insulin responses to an oral glucose challenge [22]. Most recent study found that up-regulation of GIP production through interaction with GIP receptors on islets may be a key factor in multi-hormonal dysregulation in T2DM [18]. Additionally, similar findings were reported in various animal models [23-25]. Our results are in accordance with many studies [26] supporting the role of GIP in an early pathophysiological step that could lead to T2DM. GIP secretion was also known to be preserved in response to OGTT or meal test in patients with T2DM [27], but the effect of GIP on insulin is blunted [28]. Collectively, increased GIP levels may lead to development of T2DM in similar way like insulin resistance.

However, GIP level in subjects with diabetes was not different from the GIP level of the subjects who remained normal during follow-up period. GIP level decrement after diabetes development is hard to explain because little is known about the molecular mechanism of GIP secretion, although several factors which are associated with GIP gene expression have been reported [29,30]. Recent experimental study with T2DM patients found that GIP has negligible effect on plasma glucose at fasting glycemia but it retains insulinotropic effects only during hyperglycemia [31]. Therefore, full GIP response data after meal is further needed to explain this result. Similar patterns are also seen in C-peptide changes during T2DM development. In the early of stages of T2DM, C-peptide levels are usually elevated compared to normal subjects; however, as the duration of T2DM becomes longer, C-peptide levels decrease [32]. ALT was associated with increase in the risk of T2DM even if it is not elevated in the subjects with diabetes [12]. Another explanation is that GIP level can be affected by anti-diabetic drugs. We could not adjust the effect of glucose lowering medication on the serum incretin level in the diabetes group due to lack of data. However, incretin mimetics were not available in the year of 2001 to 2002. Further investigations are required to clarify the causal relationships between the GIP level and diabetes development.

To the best of our knowledge, this is the first study that measured incretin levels in a large Asian community-based cohort. Moreover, this was an 8-year prospective study of the relationship between incretin hormones and diabetes development. The most significant limitation is that the samples were fasting samples, and thus it was impossible to determine the response of incretin hormones to meal stimuli. However, it seems reasonable to assume that fasting total GIP levels might be used to estimate enteroinsular axis like the homeostasis model assessment. In homeostasis model assessment, FPG and insulin levels are used to estimate insulin resistance and β -cell function [14]. Another limitation is that the samples were not preserved in DPP4 inhibitor containing tube.

In summary, elevated GIP levels were associated with increased diabetes risk in this study. After adjusting for other associated risk factors, fasting GIP levels might be a risk factor for the development of diabetes mellitus, suggesting that deterioration in the enteroinsular axis might occur before diabetes develops.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We are grateful to Bong Deok Kim and Hyun Kyu Kim, staff at the Center for Clinical Epidemiology at the Ajou University School of Medicine, for their effort in managing the Korean Health and Genome Study. This study was supported by the Samsung Medical Center Clinical Research Development Program grant #CRS-110-22-1 and National Genome Research EnM

Institute, the Korean Center for Disease Control and Prevention (contract #2001 to 2003-348-6111-221, 2004-347-6111-213 and 2005-347-2400-2440-215). The funding source had no role in the collection of the data or in the decision to submit the manuscript for publication.

REFERENCES

- Chia CW, Egan JM. Incretin-based therapies in type 2 diabetes mellitus. J Clin Endocrinol Metab 2008;93:3703-16.
- Nauck M, Stockmann F, Ebert R, Creutzfeldt W. Reduced incretin effect in type 2 (non-insulin-dependent) diabetes. Diabetologia 1986;29:46-52.
- Nauck MA, Heimesaat MM, Orskov C, Holst JJ, Ebert R, Creutzfeldt W. Preserved incretin activity of glucagon-like peptide 1 [7-36 amide] but not of synthetic human gastric inhibitory polypeptide in patients with type-2 diabetes mellitus. J Clin Invest 1993;91:301-7.
- Lee S, Yabe D, Nohtomi K, Takada M, Morita R, Seino Y, et al. Intact glucagon-like peptide-1 levels are not decreased in Japanese patients with type 2 diabetes. Endocr J 2010;57:119-26.
- Oh TJ, Kim MY, Shin JY, Lee JC, Kim S, Park KS, et al. The incretin effect in Korean subjects with normal glucose tolerance or type 2 diabetes. Clin Endocrinol (Oxf) 2014;80:221-7.
- Calanna S, Christensen M, Holst JJ, Laferrere B, Gluud LL, Vilsboll T, et al. Secretion of glucagon-like peptide-1 in patients with type 2 diabetes mellitus: systematic review and meta-analyses of clinical studies. Diabetologia 2013;56:965-72.
- Yabe D, Seino Y, Fukushima M, Seino S. β Cell dysfunction versus insulin resistance in the pathogenesis of type 2 diabetes in East Asians. Curr Diab Rep 2015;15:602.
- Rhee SY, Woo JT. The prediabetic period: review of clinical aspects. Diabetes Metab J 2011;35:107-16.
- Brambilla P, La Valle E, Falbo R, Limonta G, Signorini S, Cappellini F, et al. Normal fasting plasma glucose and risk of type 2 diabetes. Diabetes Care 2011;34:1372-4.
- Vollmer K, Holst JJ, Baller B, Ellrichmann M, Nauck MA, Schmidt WE, et al. Predictors of incretin concentrations in subjects with normal, impaired, and diabetic glucose tolerance. Diabetes 2008;57:678-87.
- Rask E, Olsson T, Soderberg S, Holst JJ, Tura A, Pacini G, et al. Insulin secretion and incretin hormones after oral glucose in non-obese subjects with impaired glucose tolerance.

Metabolism 2004;53:624-31.

- Cho NH, Jang HC, Choi SH, Kim HR, Lee HK, Chan JC, et al. Abnormal liver function test predicts type 2 diabetes: a community-based prospective study. Diabetes Care 2007; 30:2566-8.
- Report of the Expert Committee on the diagnosis and classification of diabetes mellitus. Diabetes Care 1997;20:1183-97.
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia 1985;28:412-9.
- Vilsboll T, Krarup T, Deacon CF, Madsbad S, Holst JJ. Reduced postprandial concentrations of intact biologically active glucagon-like peptide 1 in type 2 diabetic patients. Diabetes 2001;50:609-13.
- 16. Toft-Nielsen MB, Damholt MB, Madsbad S, Hilsted LM, Hughes TE, Michelsen BK, et al. Determinants of the impaired secretion of glucagon-like peptide-1 in type 2 diabetic patients. J Clin Endocrinol Metab 2001;86:3717-23.
- Smushkin G, Sathananthan A, Man CD, Zinsmeister AR, Camilleri M, Cobelli C, et al. Defects in GLP-1 response to an oral challenge do not play a significant role in the pathogenesis of prediabetes. J Clin Endocrinol Metab 2012;97:589-98.
- Chia CW, Odetunde JO, Kim W, Carlson OD, Ferrucci L, Egan JM. GIP contributes to islet trihormonal abnormalities in type 2 diabetes. J Clin Endocrinol Metab 2014;99:2477-85.
- McIntosh CH, Widenmaier S, Kim SJ. Glucose-dependent insulinotropic polypeptide signaling in pancreatic β-cells and adipocytes. J Diabetes Investig 2012;3:96-106.
- Holst JJ, Gromada J, Nauck MA. The pathogenesis of NI-DDM involves a defective expression of the GIP receptor. Diabetologia 1997;40:984-6.
- Ceperuelo-Mallafre V, Duran X, Pachon G, Roche K, Garrido-Sanchez L, Vilarrasa N, et al. Disruption of GIP/GIPR axis in human adipose tissue is linked to obesity and insulin resistance. J Clin Endocrinol Metab 2014;99:E908-19.
- 22. Saxena R, Hivert MF, Langenberg C, Tanaka T, Pankow JS, Vollenweider P, et al. Genetic variation in GIPR influences the glucose and insulin responses to an oral glucose challenge. Nat Genet 2010;42:142-8.
- 23. Lynn FC, Pamir N, Ng EH, McIntosh CH, Kieffer TJ, Pederson RA. Defective glucose-dependent insulinotropic polypeptide receptor expression in diabetic fatty Zucker

rats. Diabetes 2001;50:1004-11.

- 24. Gupta D, Peshavaria M, Monga N, Jetton TL, Leahy JL. Physiologic and pharmacologic modulation of glucose-dependent insulinotropic polypeptide (GIP) receptor expression in beta-cells by peroxisome proliferator-activated receptor (PPAR)-gamma signaling: possible mechanism for the GIP resistance in type 2 diabetes. Diabetes 2010;59:1445-50.
- 25. Yamada Y, Seino Y. Physiology of GIP: lesson from GIP receptor knockout mice. Horm Metab Res 2004;36:771-4.
- Cho YM, Merchant CE, Kieffer TJ. Targeting the glucagon receptor family for diabetes and obesity therapy. Pharmacol Ther 2012;135:247-78.
- 27. Calanna S, Christensen M, Holst JJ, Laferrere B, Gluud LL, Vilsboll T, et al. Secretion of glucose-dependent insulinotropic polypeptide in patients with type 2 diabetes: systematic review and meta-analysis of clinical studies. Diabetes Care 2013;36:3346-52.

- Fetner R, McGinty J, Russell C, Pi-Sunyer FX, Laferrere B. Incretins, diabetes, and bariatric surgery: a review. Surg Obes Relat Dis 2005;1:589-97.
- 29. Baggio LL, Drucker DJ. Biology of incretins: GLP-1 and GIP. Gastroenterology 2007;132:2131-57.
- Parker HE, Habib AM, Rogers GJ, Gribble FM, Reimann F. Nutrient-dependent secretion of glucose-dependent insulinotropic polypeptide from primary murine K cells. Diabetologia 2009;52:289-98.
- Christensen MB, Calanna S, Holst JJ, Vilsboll T, Knop FK. Glucose-dependent insulinotropic polypeptide: blood glucose stabilizing effects in patients with type 2 diabetes. J Clin Endocrinol Metab 2014;99:E418-26.
- 32. Neufeld ND, Raffel LJ, Landon C, Chen YD, Vadheim CM. Early presentation of type 2 diabetes in Mexican-American youth. Diabetes Care 1998;21:80-6.