

# RNAi Methodologies for the Functional Study of Signaling Molecules

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## Abstract

RNA interference (RNAi) was investigated with the aim of achieving gene silencing with diverse RNAi platforms that include small interfering RNA (siRNA), short hairpin RNA (shRNA) and antisense oligonucleotides (ASO). Different versions of each system were used to silence the expression of specific subunits of the heterotrimeric signal transducing G-proteins, G alpha i2 and G beta 2, in the RAW 264.7 murine macrophage cell line. The specificity of the different RNA interference (RNAi) platforms was assessed by DNA microarray analysis. Reliable RNAi methodologies against the genes of interest were then developed and applied to functional studies of signaling networks. This study demonstrates a successful knockdown of target genes and shows the potential of RNAi for use in functional studies of signaling molecules.

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## Introduction

RNA interference (RNAi) has been widely used by animal cell biologists as a gene silencing tool to study the cellular effects generated by modulating the expression of individual genes [1,2,3,4,5,6]. Expression can be reduced by introducing gene-specific double-stranded RNA (dsRNA) or single-stranded RNA (ssRNA) into a cell. Subsequently, the small RNA products generated from Droscha and/or Dicer-mediated dsRNA processing are delivered to the RNA-induced silencing complex (RISC), which is implicated in mRNA destruction and translational repression, and the RNA-induced transcriptional silencing complex (RITS), which is implicated in chromatin silencing [7,8,9].

However, difficulties remain in finding the best way to make RNAi work as a gene-specific silencing procedure for signaling studies. For example, dsRNA [10] or ssRNA [11,12,13] can stimulate innate cytokine responses in mammals. Additionally, synthetic small interfering RNAs (siRNAs) formulated in nonviral delivery vehicles can be potent inducers of interferons and inflammatory cytokines in mice and humans [14]. siRNAs can also mediate sequence-independent gene suppression and induce immune activation by signaling through toll-like receptor 3 (TLR3) [15]. A recent study showed that dsRNAs that are 21 nucleotides or longer are also involved in anti-angiogenic responses [16]. These could significantly limit the application of RNAi and result in off-target effects and immuno-stimulation associated with the nucleic acid treatments. In order to develop reliable RNAi reagents, three different platforms were tested: small interfering RNA (siRNA), antisense oligonucleotide (ASO) and short hairpin RNA (shRNA). siRNA, ASO and shRNA were

designed to knockdown the endogenous G protein alpha i2 (G $\alpha$ i2) or G protein beta 2 (G $\beta$ 2) gene in screens in RAW 264.7 murine macrophage-like cells. A series of gene expression profiling experiments and quantitative reverse transcriptase-polymerase chain reaction (QRT-PCR) analyses were carried out to assess both the specificity of the target gene knockdown and the functional studies of the targeted signaling molecules. G $\alpha$ i2 knockdown cells were tested to monitor the changes in signaling networks after treatment with different ligands, in this case lipopolysaccharide (LPS), Pam<sub>2</sub>CSK4 (Pam<sub>2</sub>) or prostaglandin E<sub>2</sub> (PGE<sub>2</sub>).

## Results

### DNA Microarray and RT-PCR verification of expression of specific genes in RAW 264.7 cells

To develop a clear picture of the nature of the genes expressed in RAW 264.7 cells, Affymetrix GeneChip screening was performed, producing duplicate array data sets. Affymetrix Mouse 430 chips A and B were used to assess the gene expression of RAW 264.7 cells, mouse bone marrow derived macrophage (BMDM) cells and the mouse macrophage cell line, J774A.1. BMDM and the cell lines have 'Present Call' gene elements ranging from 17K to 19K (data available at The Signaling Gateway: <http://www.signaling-gateway.org/data/micro/cgi-bin/micro.cgi?expt=affy>). Table 1 shows that a gene marked 'Present' in the Affymetrix GeneChip data was almost always determined to be 'Present' when measured using RT-PCR. To avoid any possible errors in the design of PCR primer sets, genes showing an 'Absent' signal in RT-PCR during the first trial were verified using 2 to 3 alternative

primer sets. Overall, approximately 5% of the transcripts found to be present in the microarray were absent according to RT-PCR, and 30–40% of absent determinations according to the microarray were detected by RT-PCR (Table S1). However, in cases in which multiple probes existed on the chip (38% of the probes), reproducible calls (P: Present or A: Absent) improve the odds of a correct result. The presence of *G $\alpha$ i2* and *G $\beta$ 2* mRNAs was verified using a DNA microarray and by a RT-PCR analysis (Table 1).

### Establishment of cells lacking *G $\alpha$ i2* and *G $\beta$ 2* proteins

G protein coupled receptors are characterized by seven transmembrane domains, and ligands stimulating these receptors are diverse and include immuno-stimulating molecules. Heterotrimeric G proteins are composed of  $\alpha$ ,  $\beta$  and  $\gamma$  subunits, with the G protein alpha subunits consisting of four families: *G $\alpha$ i/o*, *G $\alpha$ s*, *G $\alpha$ q/11* and *G $\alpha$ 12/13*. The *G $\alpha$ i* family is further characterized into *G $\alpha$ i1*, *G $\alpha$ i2* and *G $\alpha$ i3*. *G $\alpha$ i2* has been reported to inhibit the activities of types I, V, and VI adenylyl cyclase isoforms directly [17]. As *G $\alpha$ i2* appears to be involved in a complex pattern of

signaling [17,18,19,20,21,22,23,24,25,26,27], insight into its function was sought by silencing the *G $\alpha$ i2* transcript. In addition, the *G $\beta$ 2* subunits released upon activation of the heterotrimeric G protein activate specific effectors, and previous studies have shown it to be the primary beta subunit for certain *G $\alpha$ i* signaling pathways in macrophage cells [5,6]. Thus, *G $\beta$ 2* was also targeted to test the RNAi mechanisms.

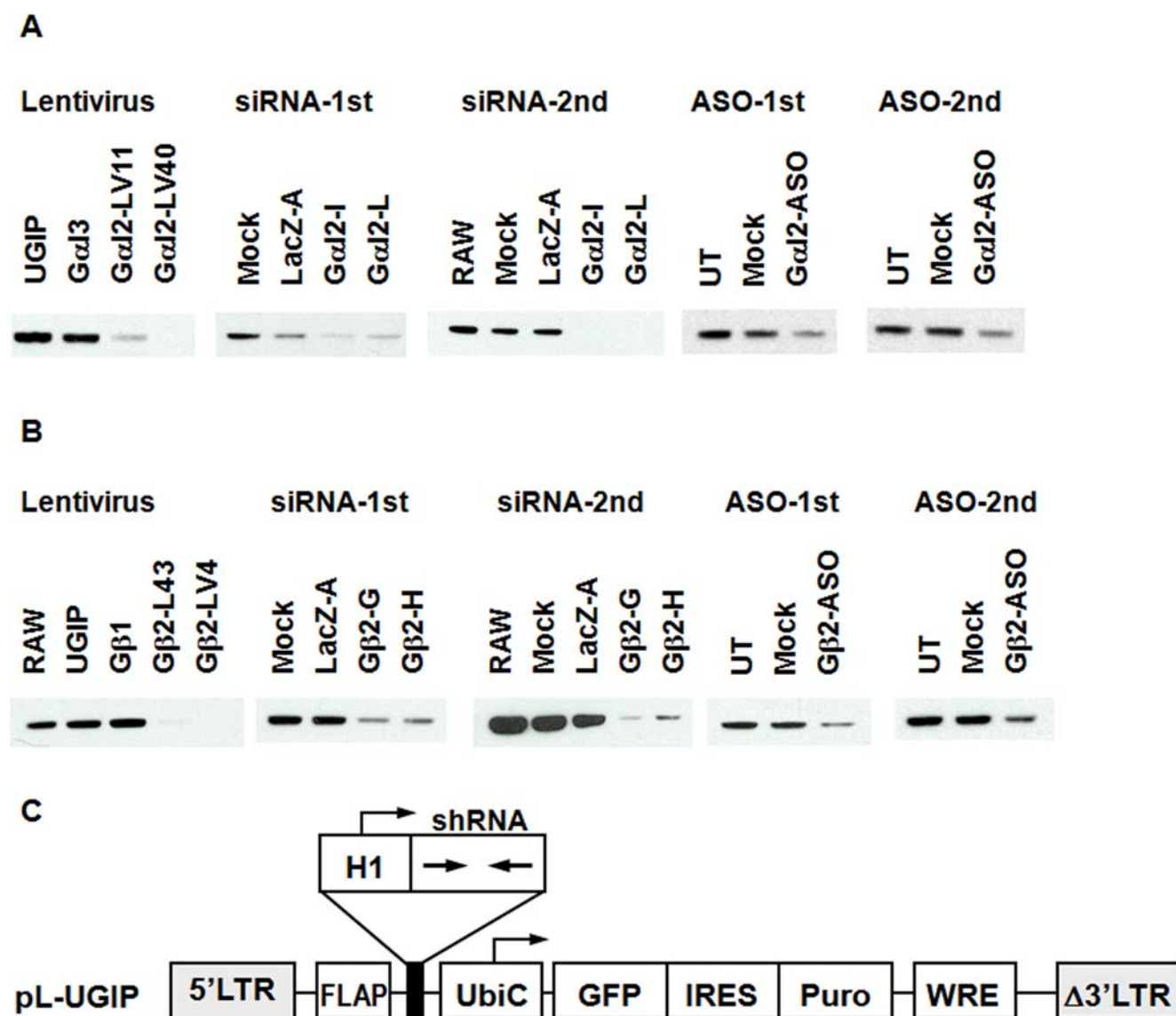
RAW 264.7 cells were transfected with siRNA or ASO against *G $\alpha$ i2* or *G $\beta$ 2*. Western blot results (Figures 1A and B) showed a significant knockdown of the target gene in each of experiments. RAW cells were transfected using FuGENE6 (Roche Molecular Biochemicals, Indianapolis, IN) for ASO and Lipofectamine 2000 (Invitrogen, Carlsbad, CA) for siRNA. Cells were harvested at 72 hr post-transfection.

Pseudotyped lentivirus carrying shRNA can be efficiently integrated into the chromosome, making it a good carrier for the delivery and sustained expression of shRNA [5,28]. Lentiviral vectors were constructed with expression cassettes that allowed the expression of antibiotic selection markers (e.g., puromycin) in order to identify and enrich the fraction of transduced cells

**Table 1.** Expression of heterotrimeric G protein subunits and RGS proteins in RAW 264.7 cells, as assessed by Affymetrix GeneChips and RT-PCR (P: present, A: absent).

Symbol	Gene Name	Affymetrix Calls	RT-PCR Results	Abundance	UMRR*
Gnai1	G alpha i1 subunit	A	A		P
Gnai2	G alpha i2 subunit	P	P	high	
Gnai3	G alpha i3 subunit	P	P	medium	
Gna11	G alpha 11 subunit	P	P	medium	
Gna14	G alpha 14 subunit	A,A	A		P
Gna15	G alpha 15/16 subunit	A	P	high	
Gnaq	G alpha q subunit	P	P	high	
Gnb1	G beta 1 subunit	P,A,P	P	high	
Gnb2	G beta 2 subunit	P	P	medium	
Gnb3	G beta 3 subunit	A	A		P
Gnb4	G beta 4 subunit	P	P	medium	
Gnb5	G beta 5 subunit	A,P	P	high	
Gng2	G gamma 2 subunit	P,A,P	P	low	
Gng3	G gamma 3 subunit	A	A		P
Gng4	G gamma 4 subunit	A,A,A	A		A
Gng5	G gamma 5 subunit	P,P,A	P	medium	
Gng7	G gamma 7 subunit	A	P	low	P
Gng8	G gamma 8 subunit	A	P	medium	
Gng10	G gamma 10 subunit	P			
Gng11	G gamma 11 subunit	A	P	medium	
Gng12	G gamma 12 subunit	P			
Gng13	G gamma 13 subunit	A	A		P
Gng14	G gamma 14 subunit	P			
Rgs4	regulator of G protein signaling 4	A,A,A	A		P
Rgs7	regulator of G protein signaling 7	A	A		P
Rgs8	regulator of G protein signaling 8		P	medium	P
Rgs13	regulator of G protein signaling 13	A	P	low	P
Rgs16	regulator of G protein signaling 16	A,A,A	P	high	
Rgs19	regulator of G protein signaling 19	P,P	P	high	P

\*RT-PCR results in Universal Mouse Reference RNA (11 cell line mixture).  
doi:10.1371/journal.pone.0004559.t001



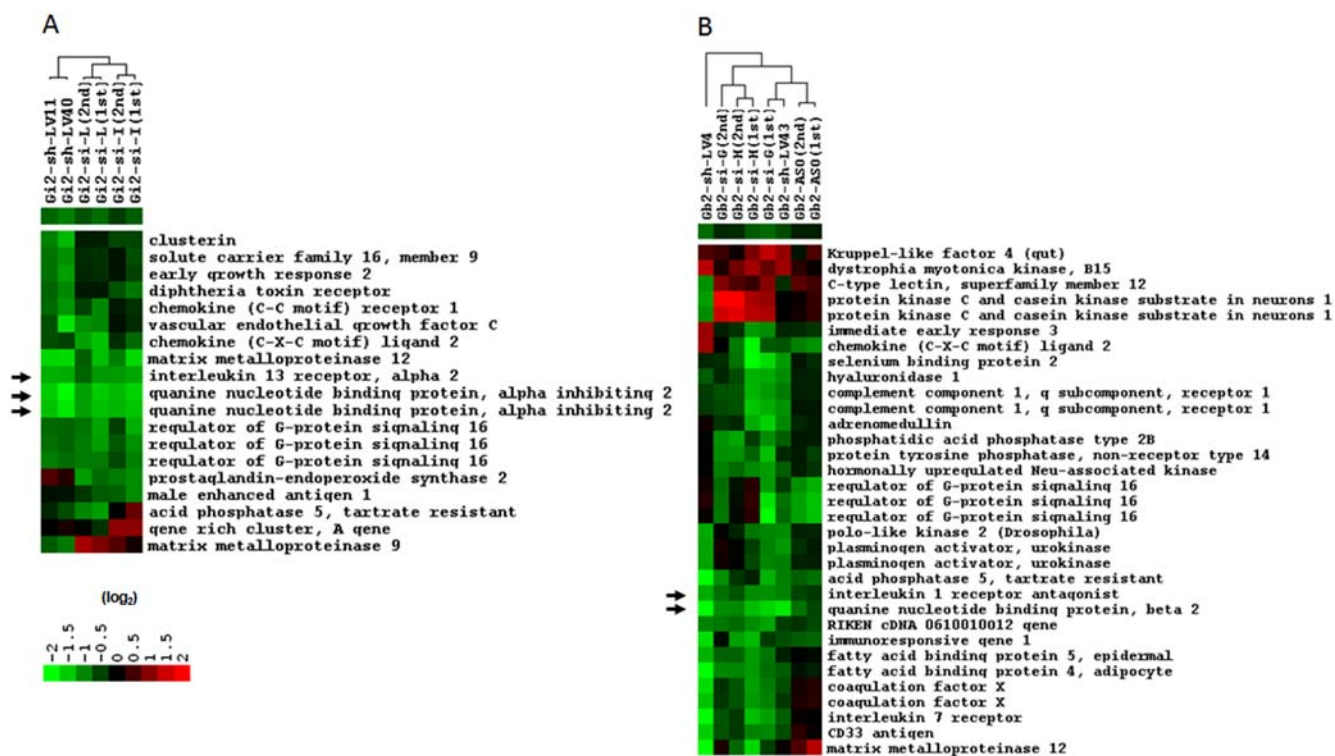
**Figure 1. Western blot analysis of (A) *Gα12* and (B) *Gβ2* proteins in shRNA, siRNA and ASO treated RAW 264.7 cells.** Two independent experiments are shown for the siRNA and ASO treatments. Blots from the shRNA-expressing lentiviral (LV) lines are representative of multiple samples taken over a 5-week period. Significant target gene knockdown is observed with all three platforms. UGIP: control lentivirus transfected cell line, UT: untreated, Mock: mock-treated. (C) Schematic diagram of the lentiviral construct used to generate shRNA expressing RAW 264.7 cell lines. The hairpin form of siRNA is expressed under the control of a mouse H1RNA polymerase III promoter. The vector also contains the enhanced GFP marker gene and the puromycin resistance gene (Puro) regulated by a UbiC promoter. IRES, internal ribosome entry site; FLAP, HIV-1 FLAP element; WRE, woodchuck hepatitis post-transcriptional regulatory element.  
doi:10.1371/journal.pone.0004559.g001

(Figure 1C). Cells were infected with a lentivirus containing a previously validated *Gα12* or *Gβ2* shRNA [5,6]. The target gene silencing activity of each shRNA in RAW264.7 cells was assessed by QRT-PCR and immunoblotting for endogenous proteins. Consistent with the QRT-PCR data, significant decreases in the levels of *Gα12* or *Gβ2* proteins were observed in the shRNA expressing cells (Figure 1).

#### siRNA, ASO and shRNA as RNAi tools

It was confirmed by DNA microarray analysis using custom-made inkjet-printed 16K oligonucleotide chips (Gene Expression Omnibus platform accession number GPL254: <http://www.ncbi.nlm.nih.gov/projects/geo/>) that the transfection of siRNA against *Gα12* or *Gβ2* showed significant levels of the target gene

knockdown (Figure 2). Cells were harvested at 48 hr post-transfection for assessment of mRNA, and gene expression was assessed using a DNA microarray and by QRT-PCR. Control samples were mock-treated with transfection reagent to determine the baseline levels of gene expression. It was found that 400 nM siRNA is invariably more effective than 100 nM or 200 nM. Although several papers have noted that siRNA reaches maximal effectiveness at a concentration lower than 100 nM [29,30], Song et al. [31] reported that primary macrophages required 1 μM siRNA to achieve a maximal effect. For any given siRNA, a comparable level of knockdown can be achieved with at least a 10-fold lower siRNA concentration using NIH3T3 cells (data not shown). The present results suggest that the requirement for a high siRNA



**Figure 2. Hierarchically clustered dendrograms of gene expression changes.** Clustering was achieved using the CLUSTER and TREEVIEW programs (<http://rana.lbl.gov/EisenSoftware.htm>). Each row represents a gene, and each column represents a particular sample. (A) *Gxi2* gene knockdown (LV40 and LV11: shRNA-expressing lentivirus transfected cell lines; I and L: I or L form of siRNA transfected into cells). (B) *Gb2* gene knockdown (LV4 and LV43: shRNA-expressing lentivirus transfected cell lines; G and H: G or H form of siRNA transfected into cells; Gb2-ASO: antisense oligonucleotide). The expression level relative to that of the control cells is provided by colors shown in log<sub>2</sub> scale. The target genes, *Gxi2* and *Gb2*, are highlighted by arrows. The apparent down-regulation of *IL13ra2* in the *Gxi2*-deficient cells and *IL1rn* in the *Gb2*-deficient cells are also highlighted (see context).

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concentration in RAW264.7 cells is simply a function of their low transfection efficiency.

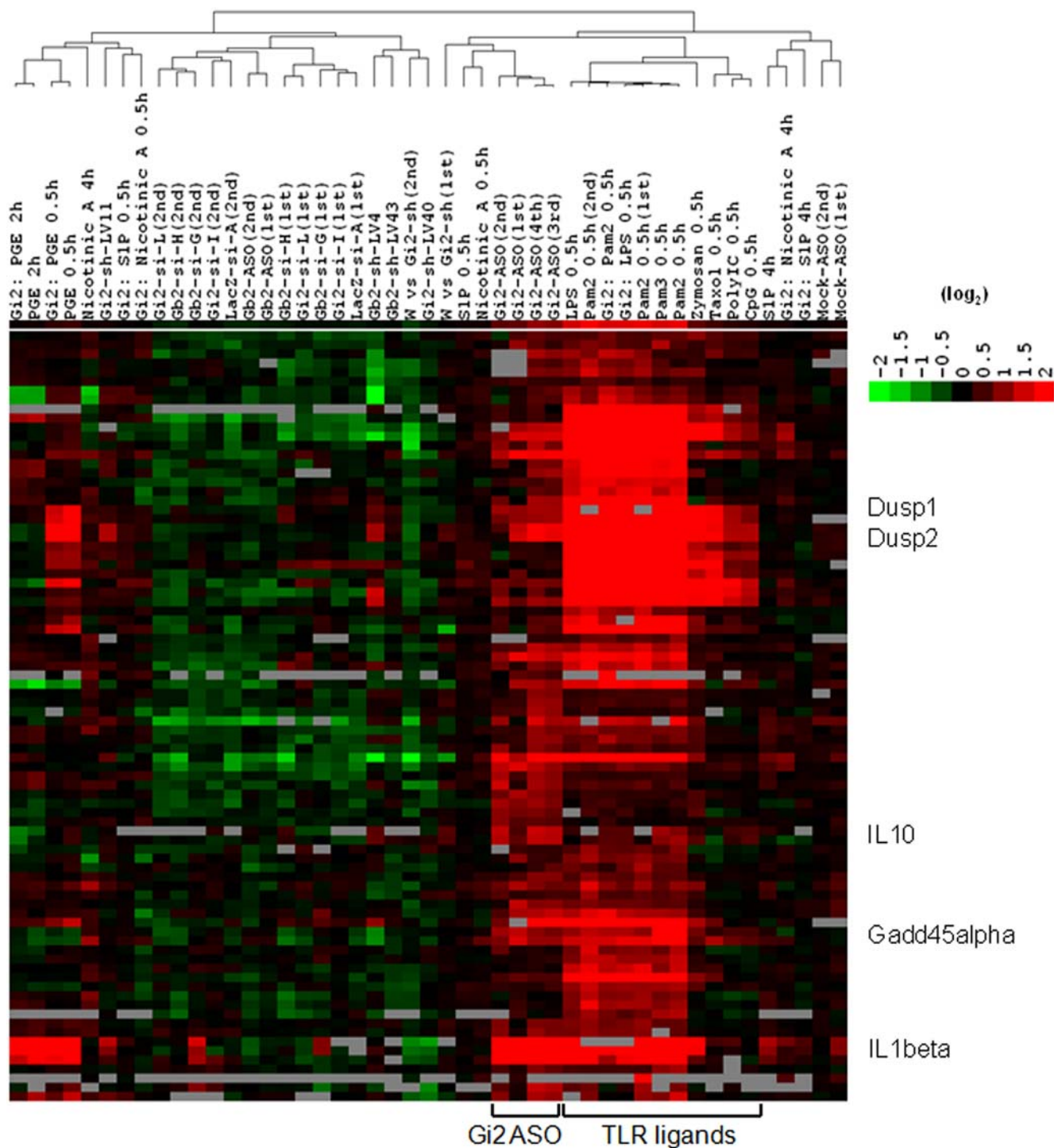
Validated ASOs against the *Gxi2* or *Gb2* gene were obtained from ISIS Pharmaceuticals, Inc. (Carlsbad, CA). The ASOs used were phosphorothioate oligodeoxynucleotides with 2'-O-methoxyethyl incorporated to enhance their affinity for RNA sequences and their resistance to degradation by nucleases. RAW 264.7 cells were transfected with 400 nM of ASO and were harvested for mRNA analysis at 48 hr post-transfection. Results from the DNA microarrays showed a detectable knockdown for ASO against *Gb2* message (Figure 2B) as well as a specific knockdown for ASO targeting *Gxi2*. Another set of *Gxi2* ASO data was not included in the analysis shown in Figure 2A due to the unexpected induction of TLR-related genes. This will be discussed later.

It has been shown that the knockdown effect mediated by siRNA or ASO is sustained for up to several days [29,32,33]. However, creation of a cell line infected with a shRNA-expressing lentivirus provides an approach for gene knockdown over a longer term. The analysis in Figure 2 shows that lentiviral vector-based RNAi works well in RAW 264.7 cells with a robust knockdown of the *Gxi2* and *Gb2* target genes.

### Induction of TLR-related genes by ASO designed for a knockdown of *Gxi2*

The ASO tests showed an unexpected observation of a significant degree of induction of TLR-related genes by one of ASOs designed for the knockdown of the *Gxi2* gene. Kim et al. [34] found a very potent induction of interferon  $\alpha$  and  $\beta$  by short

single-stranded RNAs (ssRNAs) transcribed with T3, T7 and Sp6 RNA polymerases. In their studies, analyses of the potential mediators of this response revealed that the initiating 5' triphosphate is required for interferon induction. Moreover, single-strand RNA bearing 5' phosphate could activate RIG-1 mediated anti-viral responses [35] and it was also shown that 5' triphosphate is the ligand for RIG-1 [36]. However, with the present ASO, neither RNA polymerase nor 5' triphosphate was used. Nonetheless, a transcription profiling analysis revealed patterns of regulation that were very similar to the patterns of expression induced by ligands that trigger TLRs and clustered with them (Figure 3), showing a higher expression of *IL1 $\beta$* , *IFN $\alpha$ -inducible protein*, *chemokine (C-C motif) ligand 2 (Ccl2)*, *IFN-induced protein with tetratricopeptide repeats 1 (Ifit1)*, 2 (*Ifit2*) and 3 (*Ifit3*), *granulocyte colony stimulating factor 3 (Csf3)*, *immunoresponsive gene 1 (Irg1)*, *IL6*, *guanylate nucleotide binding protein 1 (Gbp1)*, 2 (*Gbp2*) and 4 (*Gbp4*), *dual specificity phosphatase 1 (Dusp1)* and 2 (*Dusp2*), *IL10*, and *growth arrest and DNA-damage-inducible 45 $\alpha$  (Gadd45 $\alpha$ )* (not all data shown: separate paper in preparation). When the ASO sequence for *Gxi2* was studied further, it was found that the *Gxi2* antisense oligonucleotide sequence has several CpG motifs (5'-TTT AGA GCG CTC GGC TGC CG-3': multiple unmethylated CpG appeared and one purine-purine-CpG-pyrimidine-pyrimidine existed). Unmethylated CpG motifs are present in bacterial genomic DNA and function as a pattern recognition motif by the host innate immune system [37,38]. For mouse, the sequence GACGTT appears to be optimal, but flanking nucleotides to the



**Figure 3. Hierarchical clustering of gene expression for different RNAi platforms or ligand treatments in *Gai2* knockdown RAW 264.7 cells.** While Pam<sub>2</sub> 0.5 h (1st) and Pam<sub>2</sub> 0.5 h (2nd) were tested in lentivirus vector transfected RAW cell lines, Pam<sub>2</sub> 0.5 h was carried out in wild type RAW 264.7 cells together with other TLR ligand experiments, in this case Pam<sub>3</sub>, Zymosan, Taxol, PolyIC and CpG.  
doi:10.1371/journal.pone.0004559.g003

central CpG core appear to influence recognition to some degree. Oligonucleotides containing the core CpG motif might bind to the TLR9 whose expression was confirmed in RAW 264.7 cells (Table 2). TLR9 engagement triggers alteration of the cellular redox balance, tyrosine phosphorylation of vav1 by a src-related tyrosine kinase [39], and the induction of cell signaling pathways including mitogen-activated protein kinases (MAPKs)

and NFκB [37]. Interleukin-1 receptor-associated kinase (IRAK) and/or tumor-necrosis-factor-receptor-associated factor 6 (TRAF6) may be diverging points for NFκB activation in response to CpG DNA in RAW264.7 cells [40,41]. In our ASO sequence against *Gai2*, three copies of the unmethylated CpG motif per oligonucleotide led to the enhanced activation of TLRs.

**Table 2.** RT-PCR confirmation of TLR expression in RAW 264.7 cells (P: present, A: absent).

Gene Name	RT-PCR Results in RAW	Abundance	Affymetrix Chip Call
TLR1	P	High	P
TLR2	P	High	P
TLR3	P	High	P
TLR4	P	Low	P
TLR5	A		A
TLR6	P	Medium	P
TLR7	P	High	P
TLR8	A		A
TLR9	P	High	A
TLR10 (not in mouse)			
TLR11	A		
TLR12	P	Medium	
TLR13	P	High	

doi:10.1371/journal.pone.0004559.t002

### The effects of G $\alpha$ i2 or G $\beta$ 2 knockdown

Gene expression analyses were performed for all of the transduced cells using custom-made 16K oligonucleotide DNA microarrays. It is well known that the individual members of the G protein heterotrimers are necessary for the stability of their binding partners [5,42]. While the regulated number of genes in each knockdown experiment in which a two-fold cut-off was used was between about 30 to 80, changes in the mRNA expression levels of other  $\alpha$ ,  $\beta$  or  $\gamma$  G proteins were not detected in siRNA- or ASO-treated cells or in shRNA-containing cells (Table S2), indicating that the absence of the G $\alpha$ i2 or G $\beta$ 2 subunit protein did not affect the levels of transcripts encoding other individual members of the G protein heterotrimers.

In addition to the knockdown of the intended target, apparent regulation of other genes depending on the RNAi platforms was found (Figure 2). As shown in Figure 2, there appeared to be up- or down-regulations of some genes in both G $\alpha$ i2- and G $\beta$ 2-deficient cell lines. The apparent down-regulation of interleukin 13 receptor alpha2 (IL13ra2) in the G $\alpha$ i2-deficient cells and interleukin 1 receptor antagonist (IL1rn) in the G $\beta$ 2-deficient cells may be noteworthy, as these changes were observed with all platforms (Figure 2). This result is intriguing given the fact that these proteins are not reported as being capable of coupling to either G $\alpha$ i2 or G $\beta$ 2. A biological link was not found for G $\alpha$ i2 or G $\beta$ 2 to IL13ra2 or IL1rn in published reports; however, it was noted that the selected oligonucleotide sequences used for G $\alpha$ i2 or G $\beta$ 2 RNAi tests had homology of 7 to 8 contiguous nucleotides to either IL13ra2 (siRNA structure I) or IL1rn (siRNA structure G and ASO) (Figures 4A and B). While the direct silencing of nontargeted genes containing as few as eleven contiguous nucleotides of identity to the siRNA has been reported [43], it appears to be unlikely that 7 to 8 contiguous nucleotides of identity can regulate the expression of unintended targets. Furthermore, given the fact that not all of the siRNA, shRNA or ASO sequences showed significant homology to the IL13ra2 or IL1rn sequences (Figure 4), these common down-regulations may be due to an unappreciated biological consequence of G $\alpha$ i2 or G $\beta$ 2 depletion and shed light onto the unidentified functions of these G-protein subunits. Interestingly, the expression of RGS16 (Regulators of G

protein Signaling 16) was also downregulated in many of the tests for either G $\alpha$ i2 or G $\beta$ 2 (Figure 2). While RGS17 has been reported to act as a GTPase-activating protein (GAP) on free G $\alpha$ i2 and G $\alpha$ o under pre-steady-state conditions [44], it remains to be determined if the depletion of RGS16 in the G $\alpha$ i2 and G $\beta$ 2 knockdown cells is biologically relevant.

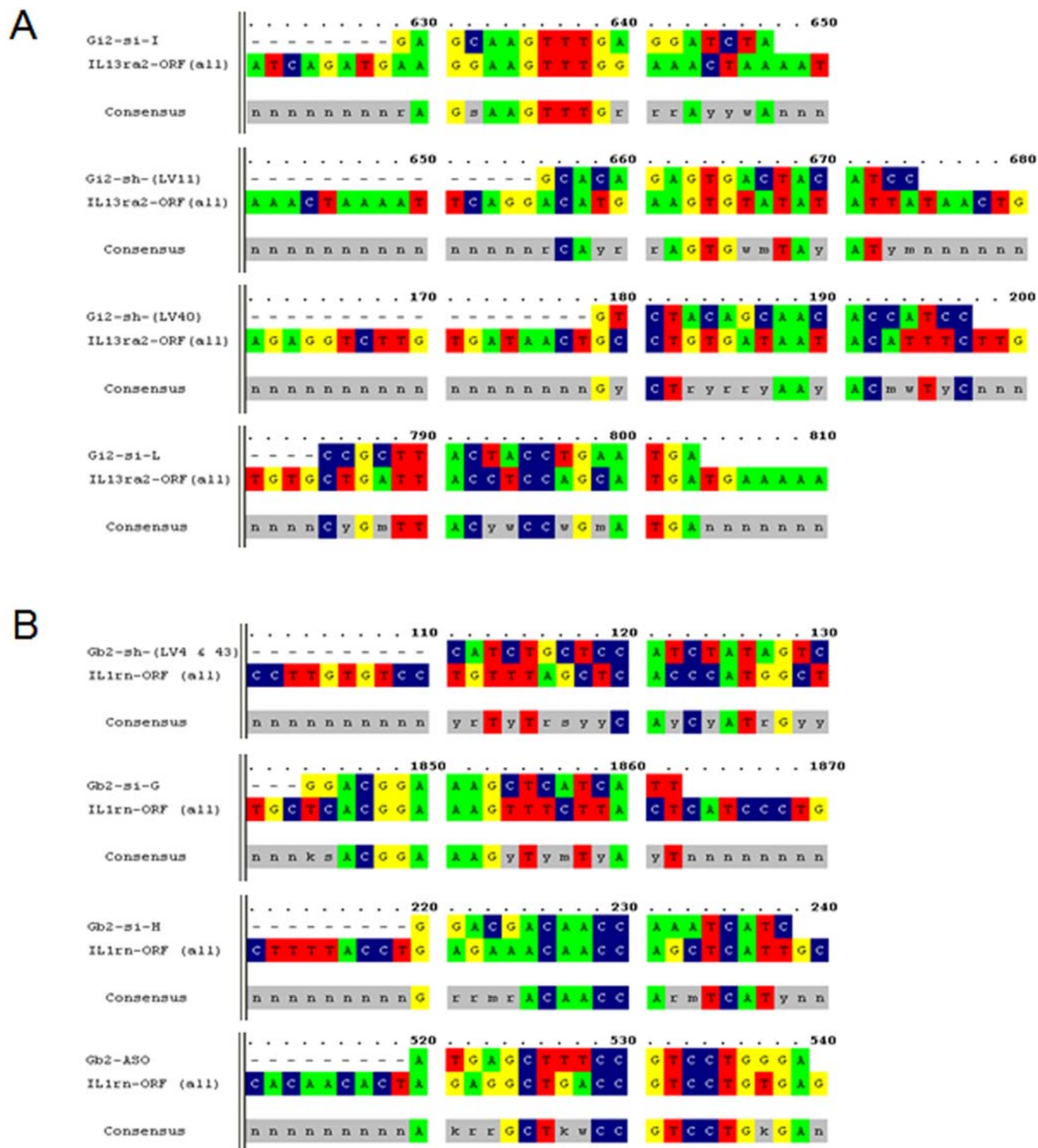
### RNAi-based perturbations

The present studies were extended to reveal which signaling networks are associated with the reduction of the G $\alpha$ i2 protein in the presence of lipopolysaccharide (LPS), Pam2CSK4 (Pam<sub>2</sub>) or prostaglandin E2 (PGE<sub>2</sub>). To observe the effect of the absence of G $\alpha$ i2 in a ligand-dependent manner, gene expression was examined in both control RAW 264.7 cells (control lentiviral vector transfected cell lines) and G $\alpha$ i2-deficient cells (G $\alpha$ i2 shRNA-harboring lentiviral cell lines) after stimulation with LPS (100 ng/ml LPS and 100 pM LPS-binding peptide), Pam<sub>2</sub> (350 nM) or PGE<sub>2</sub> (10  $\mu$ M) using 16K oligonucleotide microarrays. According to the criterion of a  $\geq 2$  fold change in expression, exposure of the cells to LPS for 30 min resulted in the up-regulation of nearly 100 transcripts in both control and G $\alpha$ i2 knockdown cells in a similar pattern. A partial image of the set of clustered dendrograms is shown in Figure 3. Pam<sub>2</sub> or PGE<sub>2</sub> treatment of both cells also led to a very similar pattern of gene expression. However, there were some subtle differences in the changes in transcript levels induced by LPS or Pam<sub>2</sub> treatment in the cells lacking the G $\alpha$ i2 protein (Table 3). These genes include early growth response 2 (Egr2), the nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, zeta (Nfkbiz), tumor necrosis factor (TNF) and the 1810011O10 RIKEN clone, whose expressions were enhanced by LPS or Pam<sub>2</sub> in G $\alpha$ i2-deficient cells.

LPS is the major constituent of the outer membrane of Gram-negative bacteria. LPS binds to the cell surface receptor CD14, which enhances TLR4-dependent LPS recognition [45,46,47]. TLR4 activation engages a set of MyD88 (myeloid differentiation primary-response protein 88) adaptor family members, including MyD88, TIRAP (TIR domain containing adaptor protein), TRIF (TIR domain containing adaptor protein inducing IFN $\beta$ ), and TRAM (TRIF related adaptor molecule). Pam<sub>2</sub> is a synthetic diacylated lipopeptide, and these lipid-modified proteins are present in the cell membranes of bacterial cell walls. The intracytoplasmic signaling events associated with Pam<sub>2</sub> stimulation are mainly engaged in TLR2/6 activation, which also leads to stimulation of the MyD88-dependent pathway [48]. Therefore, the synergistic effects of G $\alpha$ i2 knockdown on LPS- or Pam<sub>2</sub>-induced Egr2, Nfkbiz or TNF (as shown in Table 3) suggest that some aspect of these signaling pathways is influenced by G $\alpha$ i2 levels. This may reflect a heightened response or defective negative regulation in G $\alpha$ i2-deficient cells. Consistent with the present results, stimulation of G $\alpha$ i2-deficient peripheral T cells induced a hyper-responsive profile of interleukin-2, tumor necrosis factor, and interferon-gamma production [18]. Unlike G $\alpha$ i1 or G $\alpha$ i3, cells deficient in G $\alpha$ i2 were reported to be hyper-responsive to cytokines, including IFN-gamma and IL-4 production following activation [20].

### Discussion

It is well established that siRNA efficacy is determined by how effectively the siRNA is incorporated into RISC [49,50,51]. This necessitates the testing of several candidate siRNAs to identify an effective reagent. Recently, it has been shown that testing of many siRNAs can be circumvented by *in vitro* digestion of a long dsRNA



**Figure 4. Sequence homologies of the oligonucleotides used in siRNA, shRNA or ASO for (A) *Gai2* or (B) *Gβ2*.** Sequences were analyzed by OMIGA 2.0 (Rainbow Technologies, Inc.). doi:10.1371/journal.pone.0004559.g004

using the dicer enzyme. This produces a population of siRNAs covering a much larger region of target sequence and usually obviates the need to test multiple sequences for each gene. The caveats, however, are that this approach theoretically increases the possibility of an undesirable knockdown of other genes with significant homology to the target sequences used as the dicer template, and

siRNAs derived from synthesized long dsRNAs by the T7 RNA polymerase system can trigger a potent induction of interferon  $\alpha$  and  $\beta$  in a variety of cell lines due to the initiating 5' triphosphate [34].

Three different silencing/delivery systems (siRNA, ASO and shRNA) and different sequences in the target genes in each case were used to assess how the knockdown of a particular gene may

**Table 3.** Ligand-induced gene expression changes in *G $\alpha$ i2*-deficient cells.

Gene	Assay	W vs Gi2-sh	LPS 0.5 h	Gi2: LPS 0.5 h	F: LPS	Pam2 0.5 h	Gi2: Pam2 0.5 h	F: Pam2	PGE 0.5 h	Gi2: PGE 0.5 h	F: PGE
Nfkbiz	Microarray	1.03	24.94	30.70	<b>5.76</b>	27.80	30.05	<b>2.25</b>	1.41	1.34	-0.07
	QRT-PCR		72.78	123.26	<b>50.48</b>	86.74	131.52	<b>44.79</b>	1.48	1.34	-0.14
Egr2	Microarray	0.60	12.11	18.10	<b>5.99</b>	14.80	20.74	<b>5.94</b>	0.71	0.78	0.14
	QRT-PCR		21.17	25.23	<b>4.06</b>	16.86	45.33	<b>28.47</b>	0.76	0.52	-0.62
1810011010Rik	Microarray	1.01	4.81	10.16	<b>5.35</b>	4.00	8.63	<b>4.63</b>	1.42	1.34	-0.09
	QRT-PCR		53.51	84.18	<b>30.67</b>	71.06	116.53	<b>45.47</b>	3.45	2.32	-1.13
Tnf	Microarray	0.92	7.99	13.17	<b>5.18</b>	7.93	18.22	<b>10.29</b>	0.69	0.66	-0.06
	QRT-PCR		15.44	23.70	<b>8.26</b>	15.86	20.93	<b>5.07</b>	0.92	0.55	-0.73

Note: The numbers are in fold differences. F (F factor) was calculated by (ligand effect in knockdown) - (ligand effect in wild type): Ligand effect =  $2^X - 1$ , if  $X \geq 0$  or  $= 1 - 1/2^X$ , if  $X < 0$ .

Where, X is the  $\log_2$  ratio.

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affect molecular mechanisms. The significant knockdown in the *G $\alpha$ i2* and *G $\beta$ 2* protein levels suggests that the knockdown levels achievable with siRNA, ASO or shRNA may be sufficient to assess a transient or permanent phenotype [29].

'Off-target effects' can occur if the sequence identity between the siRNA and random mRNA transcripts is high enough to cause RNAi to knockdown the expression of non-targeted genes. However, in our experiments, sequence-specific off-target effects were not obvious except for the ASO designed for *G $\alpha$ i2*, which indicates that the design of oligonucleotides is certainly important and should be tested in the genomic scale before it is used to knockdown a target gene in biological studies. Kim et al. [29] and Siolas et al. [52] found that synthetic RNA duplexes ~27 nucleotides in length can be up to 100 times more potent than traditional 21-unit siRNA oligomers. Enhancing the potency of RNAi duplexes and thus lowering the effective concentration of that molecule is another preferable working direction to minimize off-target effects. The enhanced potency of the ~27 nucleotide siRNA is attributed to the fact that it is diced by the dicer and directly delivered to the RNA-induced silencing complex (RISC).

Extracellular stimulants induce various cellular responses by regulating diverse pathways, and perturbation of any genes in the pathway can generate altered regulation of signaling networks. In a mouse model, *in vitro* stimulation of splenocytes with formalin-killed *Staphylococcus aureus* resulted in significantly increased production of IL1 $\beta$ , TNF, and IL12p40 in *G $\alpha$ i2* (-/-) compared to control mice [26]. Mice with targeted deletion of *G $\alpha$ i2* develop an inflammatory bowel disease closely resembling ulcerative colitis, and the IFN $\gamma$  and IL1 $\beta$  levels were increased in the inflamed colons [53]. In RAW 264.7 cells, the expression level of *G $\alpha$ i2* was relatively high compared to that of *G $\alpha$ i3*, whereas *G $\alpha$ i1* was not expressed (Table 1). *G $\alpha$ i2* proteins may play a role in modulating LPS-activated signaling through TLR4, leading to inflammatory mediator production in RAW 264.7 cells. There was a significant increase in the LPS-induced production of TNF $\alpha$  in *G $\alpha$ i2*-deficient RAW 264.7 cells compared with control cells (Table 3). Subsequent QRT-PCR studies also confirmed that TNF $\alpha$  levels following a LPS challenge were significantly greater in *G $\alpha$ i2*-deficient cells. This is consistent with the published data from *G $\alpha$ i2* (-/-) mice [26,54].

Taken together, success was obtained with the knockdown of target genes in RAW 264.7 cells using chemically synthesized siRNA, lentiviral shRNA and ASO. The specificity of the different platforms of RNAi was also assessed through DNA microarrays,

and the findings showed the successful development of siRNA reagents against the genes of interest and the useful application of vector-based RNAi to RAW 264.7 cells for functional studies. Well-designed RNAi would be a very useful tool to knockdown specific target genes and to study the functional roles of molecules within the relevant signaling network system.

## Materials and Methods

### Nucleic acid sequences for shRNAs, siRNAs and ASOs

The shRNAs, siRNAs and ASOs used in these studies were based on the following sequences: shRNAs (*G $\alpha$ i2*-LV40: 5'-GTC TAC AGC AAC ACC ATC C-3', *G $\alpha$ i2*-LV11: 5'-GCA CAG AGT GAC TAC ATC C-3', *G $\beta$ 2*-LV43 & 4: 5'-CAT CTG CTC CAT CTA TAG TC-3'), siRNAs (*G $\alpha$ i2*-I: 5'-GAG CAA GTT TGA GGA TCT A-3', *G $\alpha$ i2*-L: 5'-CCG CTT ACT ACC TGA ATG A-3', *G $\beta$ 2*-G: 5'-GGA CGG AAA GCT CAT CAT T-3', *G $\beta$ 2*-H: 5'-GGA CGA CAA CCA AAT CAT C-3') and ASOs (*G $\alpha$ i2*: 5'-TTT AGA GCG CTC GGC TGC CG-3', *G $\beta$ 2*: 5'-ATG AGC TTT CCG TCC TGG GA-3').

### siRNAs or ASO treatments

All ASOs in this study were synthesized by ISIS Pharmaceuticals, and all siRNAs were synthesized and HPLC-purified by Qiagen Inc. (Valencia, CA). Briefly, RAW 264.7 cells in 24-well dishes were transfected with 400 nM of either ASO or siRNA using either FuGENE6 (Roche Molecular Biochemicals, Indianapolis, IN) or Lipofectamine 2000 (Invitrogen, Carlsbad, CA), respectively. For siRNA transfection, an incubation volume of 200  $\mu$ l was used for 4 hr followed by the addition of media up to 1 ml. For ASO transfection, an incubation volume of 500  $\mu$ l was used. siRNA transfections were repeated 24 hr after the initial transfection while a single transfection was done for ASO treatments. Cells were harvested at 48 hr post-transfection for isolation of the RNA or at 72 hr post-transfection for isolation of the protein.

### Virus construction

Lentiviral vectors were constructed and used for virus generation as described previously [5,6]. Annealed shRNA linkers were ligated into a *Bam*H1/*Xho*1-digested pEN\_mH1c plasmid. The cassette containing the mH1 promoter and the shRNA was subcloned to a lentiviral expression vector by site-specific recombination using the Gateway system (Invitrogen). The



lentiviral vector contains the GFP gene driven by the ubiquitin promoter followed by an IRES (internal ribosome entry site) sequence and an antibiotic resistance gene for the selection of infected cells. The resultant lentiviral vectors were transfected into 293T cells two other plasmids referred to as ‘packaging’ and ‘envelope’ plasmids: pCMVΔR8.91 (expresses HIV *gag*, *pol* and *rev* genes), and pMD.G (expresses vesicular stomatitis virus G envelope protein), respectively. The packaging and envelope plasmids were a gift from the Didier Trono Lab, Geneva, Switzerland [55]. At 48 hr post-transfection, culture supernatants were collected and concentrated using Centricon Plus-80 units according to the manufacturer’s instructions (Millipore Corporate, Billerica, MA). Virus was titrated by infecting 293T cells and assessing the percent of GFP positive cells using cytometric analysis 48 hr post-infection. The titered virus was used to infect RAW 264.7 cells at a MOI (multiplicity of infection) of five 293T-transducing units per RAW 264.7 cell, at virus concentrations of 1 to  $5 \times 10^7$ /ml. These infection conditions routinely resulted in an average of 20% transduction efficiency, suggesting that a single productive viral integration occurs for every 25 viruses used on RAW 264.7 cells. Infected cells were selected according to their antibiotic resistance. These infection conditions routinely resulted in 10–25% transduction efficiency, suggesting that approximately one productive viral integration occurs for every 5 viruses used on RAW 264.7 cells. Infected cells were selected according to their antibiotic resistance.

### Western blotting

Cells were lysed with lysis buffer containing 150 mM NaCl, 50 mM NaF, 20 mM Tris, pH 7.5, 1% Triton X-100, 0.5% sodium deoxycholate, 0.1% SDS, and protease inhibitor mixture (Roche Diagnostics).

Thirty  $\mu$ g of protein was run on a NuPAGE 4–12% Bis-Tris gel (Invitrogen). Total protein was transferred to a nitrocellulose membrane (Schleicher and Schuell BioScience, Inc., Keene, NH) and was subsequently immunoblotted for the protein of interest. The membrane was blocked overnight at 4°C, which was followed by incubation with protein specific primary antibody for 2 to 3 hr at room temperature. After incubation with a secondary antibody conjugated to HRP (horseradish peroxidase) (Amersham Biosciences, Piscataway, NJ) for 1 hr at room temperature, the membrane was developed using a SuperSignal West Pico Chemiluminescent Substrate (Pierce Biotechnology, Inc., Rockford, IL) and exposed to X-OMAT film (Kodak).

### DNA microarray

Cells were cultured in media containing 0.5% FBS (for PGE<sub>2</sub>, S1P and nicotinic acid) or 10% FBS (for LPS, Pam<sub>2</sub>, Pam<sub>3</sub>, CpG, zymosan and polyIC) for 18 hr; stimulated with an agonist for 30 min, 2 hr or 4 hr; and were then harvested with Trizol (Invitrogen, Carlsbad, CA). Cells for the basic RNAi tool tests of

siRNA, shRNA or ASO were cultured in media containing 10% FBS and were harvested with Trizol. Three micrograms of total RNAs from the cells were used as the starting material for the microarray analysis.

The 16K mouse oligonucleotide arrays were fabricated using an inkjet-printing method by Agilent Technologies (Palo Alto, CA). These oligo arrays includes 13,536 probes of 70mers (Operon Technologies Inc., Alameda, CA) and 2,304 probes of 65mers (Sigma-Genosys, The Woodlands, TX). The platform description (platform accession number GPL254) is available at GEO (Gene Expression Omnibus: <http://www.ncbi.nlm.nih.gov/projects/geo/>). The aminoalyl method was utilized for the preparation of the fluorescently labeled target samples.

### Quantitative RT-PCR

Quantitative real-time RT-PCR was performed using iCycler (Bio-Rad Laboratories, Inc., Hercules, CA) following the manufacturer’s protocol. The measurement was normalized to a  $\beta$  actin RNA control. The primers of genes are as follows: Nfkbiz, F 5'-CGA TGG ACC GGT TTG CA-3', R 5'-GTA GGC GTT TGC GGT GAT G-3'; Egr-2, F 5'-GTG CCA GCT GCT ATC CAG AAG, R 5'-GGC TGT GGT TGA AGC TGG AG-3'; TNF, F 5'-CCC TCA CAC TCA GAT CAT CTT CT-3', R 5'-GCT ACG ACG TGG GCT ACA G-3';  $\beta$ -actin, F 5'-CTT TGC AGC TCC TTC GTT GC-3', R 5'-ACG ATG GAG GGG AAT ACA GC-3'. The remaining primer sequences are available upon request.

### Supporting Information

**Table S1** Expression of selected signaling genes in RAW 264.7 cells, as assessed by Affymetrix GeneChips and RT-PCR. There were multiple probe sets in some cases. U: unavailable, P: present, A: absent, M: marginal.

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**Table S2** mRNA expression levels ( $\log_2$  ratio) of G protein subunits assessed by DNA microarrays in shRNA, siRNA and ASO-treated RAW 264.7 cells.

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### Author Contributions

Conceived and designed the experiments: GL SC. Performed the experiments: LAS MSC. Wrote the paper: GL SC.

### References

- Guo S, Kemphues KJ (1995) par-1, a gene required for establishing polarity in *C. elegans* embryos, encodes a putative Ser/Thr kinase that is asymmetrically distributed. *Cell* 81: 611–620.
- Hannon GJ (2002) RNA interference. *Nature* 418: 244–251.
- Hutvagner G, Zamore PD (2002) RNAi: nature abhors a double-strand. *Current Opinion in Genetics & Development* 12: 225–232.
- Sharp PA (1999) RNAi and double-strand RNA. *Genes & Development* 13: 139–141.
- Hwang JI, Fraser ID, Choi S, Qin XF, Simon MI (2004) Analysis of C5a-mediated chemotaxis by lentiviral delivery of small interfering RNA. *Proc Natl Acad Sci U S A* 101: 488–493.
- Hwang JI, Choi S, Fraser ID, Chang MS, Simon MI (2005) Silencing the expression of multiple G $\beta$ -subunits eliminates signaling mediated by all four families of G proteins. *Proc Natl Acad Sci U S A*.
- Mello CC, Conte D Jr (2004) Revealing the world of RNA interference. *Nature* 431: 338–342.
- Bernstein E, Caudy AA, Hammond SM, Hannon GJ (2001) Role for a bidentate ribonuclease in the initiation step of RNA interference. *Nature* 409: 363–366.
- Bernstein E, Kim SY, Carmell MA, Murchison EP, Alcorn H, et al. (2003) Dicer is essential for mouse development. *Nat Genet* 35: 215–217.
- Alexopoulou L, Holt AC, Medzhitov R, Flavell RA (2001) Recognition of double-stranded RNA and activation of NF- $\kappa$ B by Toll-like receptor 3. *Nature* 413: 732–738.

11. Heil F, Hemmi H, Hochrein H, Ampenberger F, Kirschning C, et al. (2004) Species-specific recognition of single-stranded RNA via toll-like receptor 7 and 8. *Science* 303: 1526–1529.
12. Lund JM, Alexopoulou L, Sato A, Karow M, Adams NC, et al. (2004) Recognition of single-stranded RNA viruses by Toll-like receptor 7. *Proc Natl Acad Sci U S A* 101: 5598–5603.
13. Diebold SS, Kaisho T, Hemmi H, Akira S, Reis e Sousa C (2004) Innate antiviral responses by means of TLR7-mediated recognition of single-stranded RNA. *Science* 303: 1529–1531.
14. Judge AD, Sood V, Shaw JR, Fang D, McClintock K, et al. (2005) Sequence-dependent stimulation of the mammalian innate immune response by synthetic siRNA. *Nat Biotechnol* 23: 457–462.
15. Kariko K, Bhuyan P, Capodici J, Weissman D (2004) Small interfering RNAs mediate sequence-independent gene suppression and induce immune activation by signaling through toll-like receptor 3. *J Immunol* 172: 6545–6549.
16. Kleinman ME, Yamada K, Takeda A, Chandrasekaran V, Nozaki M, et al. (2008) Sequence- and target-independent angiogenesis suppression by siRNA via TLR3. *Nature* 452: 591–597.
17. Grishina G, Berlot CH (1997) Identification of common and distinct residues involved in the interaction of alpha2i and alphas with adenylyl cyclase. *Journal of Biological Chemistry* 272: 20619–20626.
18. Rudolph U, Finegold MJ, Rich SS, Harriman GR, Srinivasan Y, et al. (1995) Gi2 alpha protein deficiency: a model of inflammatory bowel disease. *Journal of Clinical Immunology* 15: 101S–105S.
19. Pace AM, Faure M, Bourne HR (1995) Gi2-mediated activation of the MAP kinase cascade. *Mol Biol Cell* 6: 1685–1695.
20. Huang TT, Zong Y, Dalwadi H, Chung C, Miceli MC, et al. (2003) TCR-mediated hyper-responsiveness of autoimmune Galphai2(-/-) mice is an intrinsic naive CD4(+) T cell disorder selective for the Galphai2 subunit. *Int Immunol* 15: 1359–1367.
21. Lyons J, Landis CA, Harsh G, Vallar L, Grunewald K, et al. (1990) Two G protein oncogenes in human endocrine tumors. *Science* 249: 655–659.
22. Sprang SR (1997) G protein mechanisms: insights from structural analysis. *Annual Review of Biochemistry* 66: 639–678.
23. Watkins DC, Johnson GL, Malbon CC (1992) Regulation of the differentiation of teratocarcinoma cells into primitive endoderm by G alpha i2. *Science* 258: 1373–1375.
24. Zhang Y, Finegold MJ, Jin Y, Wu MX (2005) Accelerated transition from the double-positive to single-positive thymocytes in G alpha i2-deficient mice. *Int Immunol* 17: 233–243.
25. Bjursten M, Hultgren Hornquist E (2005) Dietary antigen-specific T-cell responses: switch from an interleukin-10-dominated response in normal mice to a T-helper 1 cytokine profile in Galphai2-deficient mice prior to colitis. *Scandinavian Journal of Immunology* 61: 29–35.
26. Bjursten M, Hultgren OH, Hultgren Hornquist E (2004) Enhanced pro-inflammatory cytokine production in Galphai2-deficient mice on colitis prone and colitis resistant 129Sv genetic backgrounds. *Cellular Immunology* 228: 77–80.
27. Baritono E, Ceolotto G, Papparella I, Sartori M, Ciccariello L, et al. (2004) Abnormal regulation of G protein alpha(i2) subunit in skin fibroblasts from insulin-resistant hypertensive individuals. *J Hypertens* 22: 783–792.
28. Stewart SA, Dykxhoorn DM, Palliser D, Mizuno H, Yu EY, et al. (2003) Lentivirus-delivered stable gene silencing by RNAi in primary cells. *Rna-A Publication of the Rna Society* 9: 493–501.
29. Kim DH, Behlke MA, Rose SD, Chang MS, Choi S, et al. (2005) Synthetic dsRNA Dicer substrates enhance RNAi potency and efficacy. *Nat Biotechnol* 23: 222–226.
30. Overhoff M, Alken M, Far RK, Lemaitre M, Lebleu B, et al. (2005) Local RNA target structure influences siRNA efficacy: a systematic global analysis. *J Mol Biol* 348: 871–881.
31. Song E, Lee SK, Dykxhoorn DM, Novina C, Zhang D, et al. (2003) Sustained small interfering RNA-mediated human immunodeficiency virus type 1 inhibition in primary macrophages. *J Virol* 77: 7174–7181.
32. Endesfelder S, Kliche A, Lochmuller H, von Moers A, Speer A (2005) Antisense oligonucleotides and short interfering RNAs silencing the cyclin-dependent kinase inhibitor p21 improve proliferation of Duchenne muscular dystrophy patients' primary skeletal myoblasts. *J Mol Med* 83: 64–71.
33. Yin C, Xi L, Wang X, Eapen M, Kukreja RC (2005) Silencing heat shock factor 1 by small interfering RNA abrogates heat shock-induced cardioprotection against ischemia-reperfusion injury in mice. *J Mol Cell Cardiol*.
34. Kim DH, Longo M, Han Y, Lundberg P, Cantin E, et al. (2004) Interferon induction by siRNAs and ssRNAs synthesized by phage polymerase. *Nat Biotechnol* 22: 321–325.
35. Pichlmair A, Schulz O, Tan CP, Naslund TI, Liljestrom P, et al. (2006) RIG-I-mediated antiviral responses to single-stranded RNA bearing 5'-phosphates. *Science* 314: 997–1001.
36. Hornung V, Ellegast J, Kim S, Brzozka K, Jung A, et al. (2006) 5'-Triphosphate RNA is the ligand for RIG-I. *Science* 314: 994–997.
37. Krieg AM (2002) CpG motifs in bacterial DNA and their immune effects. *Annu Rev Immunol* 20: 709–760.
38. Yi AK, Yoon JG, Yeo SJ, Hong SC, English BK, et al. (2002) Role of mitogen-activated protein kinases in CpG DNA-mediated IL-10 and IL-12 production: central role of extracellular signal-regulated kinase in the negative feedback loop of the CpG DNA-mediated Th1 response. *J Immunol* 168: 4711–4720.
39. Stovall SH, Yi AK, Meals EA, Talati AJ, Godambe SA, et al. (2004) Role of vav1- and src-related tyrosine kinases in macrophage activation by CpG DNA. *J Biol Chem* 279: 13809–13816.
40. Yeo SJ, Gravis D, Yoon JG, Yi AK (2003) Myeloid differentiation factor 88-dependent transcriptional regulation of cyclooxygenase-2 expression by CpG DNA: role of NF-kappaB and p38. *J Biol Chem* 278: 22563–22573.
41. Yeo SJ, Yoon JG, Yi AK (2003) Myeloid differentiation factor 88-dependent post-transcriptional regulation of cyclooxygenase-2 expression by CpG DNA: tumor necrosis factor-alpha receptor-associated factor 6, a diverging point in the Toll-like receptor 9-signaling. *J Biol Chem* 278: 40590–40600.
42. Wang Q, Mullah BK, Robishaw JD (1999) Ribozyme approach identifies a functional association between the G protein beta1gamma7 subunits in the beta-adrenergic receptor signaling pathway. *Journal of Biological Chemistry* 274: 17365–17371.
43. Jackson AL, Bartz SR, Schelter J, Kobayashi SV, Burchard J, et al. (2003) Expression profiling reveals off-target gene regulation by RNAi. *Nature Biotechnology* 21: 635–637.
44. Mao H, Zhao Q, Daigle M, Ghahremani MH, Chidiac P, et al. (2004) RGS17/RGSZ2, a novel regulator of Gi/o, Gz, and Gq signaling. *J Biol Chem* 279: 26314–26322.
45. Akira S, Takeda K, Kaisho T (2001) Toll-like receptors: critical proteins linking innate and acquired immunity. *Nat Immunol* 2: 675–680.
46. Yamamoto M, Sato S, Hemmi H, Uematsu S, Hoshino K, et al. (2003) TRAM is specifically involved in the Toll-like receptor 4-mediated MyD88-independent signaling pathway. *Nat Immunol* 4: 1144–1150.
47. Krishnan J, Selvarajoo K, Tsuchiya M, Lee G, Choi S (2007) Toll-like receptor signal transduction. *Exp Mol Med* 39: 421–438.
48. Takeda K, Kaisho T, Akira S (2003) Toll-like receptors. *Annu Rev Immunol* 21: 335–376.
49. Castanotto D, Sakurai K, Lingeman R, Li H, Shively L, et al. (2007) Combinatorial delivery of small interfering RNAs reduces RNAi efficacy by selective incorporation into RISC. *Nucleic Acids Res* 35: 5154–5164.
50. Tafer H, Ameres SL, Obernosterer G, Gebeshuber CA, Schroeder R, et al. (2008) The impact of target site accessibility on the design of effective siRNAs. *Nat Biotechnol* 26: 578–583.
51. Koller E, Propp S, Murray H, Lima W, Bhat B, et al. (2006) Competition for RISC binding predicts in vitro potency of siRNA. *Nucleic Acids Res* 34: 4467–4476.
52. Siolas D, Lerner C, Burchard J, Ge W, Linsley PS, et al. (2005) Synthetic shRNAs as potent RNAi triggers. *Nat Biotechnol* 23: 227–231.
53. Hornquist CE, Lu X, Rogers-Fani PM, Rudolph U, Shappell S, et al. (1997) G(alpha)i2-deficient mice with colitis exhibit a local increase in memory CD4+ T cells and proinflammatory Th1-type cytokines. *Journal of Immunology* 158: 1068–1077.
54. Fan H, Zingarelli B, Peck OM, Teti G, Tempel GE, et al. (2005) Lipopolysaccharide and Gram-positive Bacteria Induced Cellular Inflammatory Responses: Role of Heterotrimeric G[alpha]i Proteins. *Am J Physiol Cell Physiol*.
55. Salmon P, Kindler V, Ducrey O, Chapuis B, Zubler RH, et al. (2000) High-level transgene expression in human hematopoietic progenitors and differentiated blood lineages after transduction with improved lentiviral vectors. *Blood* 96: 3392–3398.